

# *How to use MENAP?*

*<http://ieg4.rccc.ou.edu/mena/>*

**Version 20251202**

**Institute for Environmental Genomics (IEG)  
University of Oklahoma, Norman, OK, USA**

Daliang Ning ([ningdaliang@ou.edu](mailto:ningdaliang@ou.edu))



# Outline

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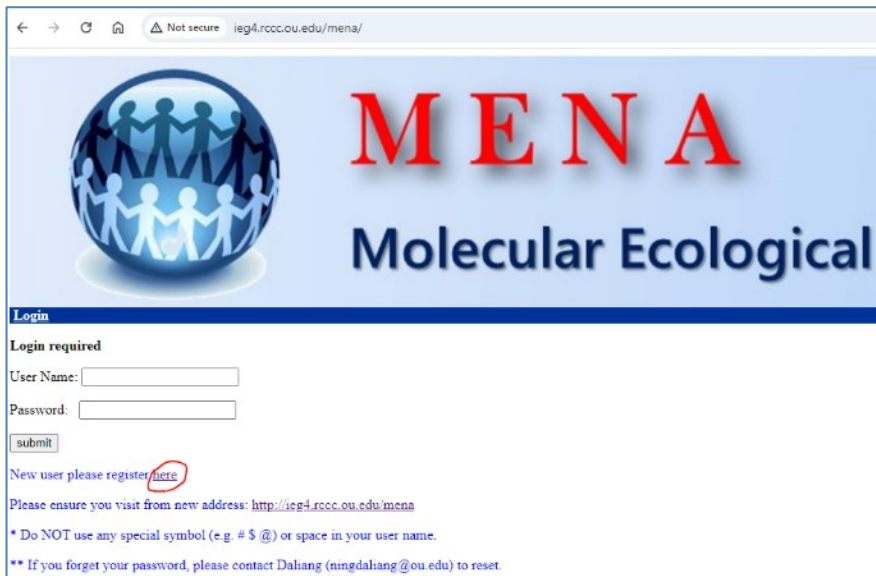
# Data requirements

- **Sample number: better  $\geq 8$** 
  - Remember fewer samples lead to less reliable correlation coefficients.
- **Taxa number: qualified taxa  $> 50$  and  $< 2000$** 
  - ‘Qualified taxa’ mean taxa detected in more than a certain number of samples
    - MENAP default setting is 50% of samples (see ‘majority’ in ‘construct network’ step)
  - **If too few, not recommend to use MENAP.**
  - **If too many, consider denoising the data or ask the maintainer for help.**


Current maintainers: Daliang Ning ([ningdaliang@ou.edu](mailto:ningdaliang@ou.edu))  
Yajiao Wang ([Yajiao.Wang-1@ou.edu](mailto:Yajiao.Wang-1@ou.edu))

# Registration

- <http://ieg4.rccc.ou.edu/mena/register.cgi>



← → ↻ 🏠 Not secure ieg4.rccc.ou.edu/mena/



# MENA

## Molecular Ecological

**Login**

Login required

User Name:

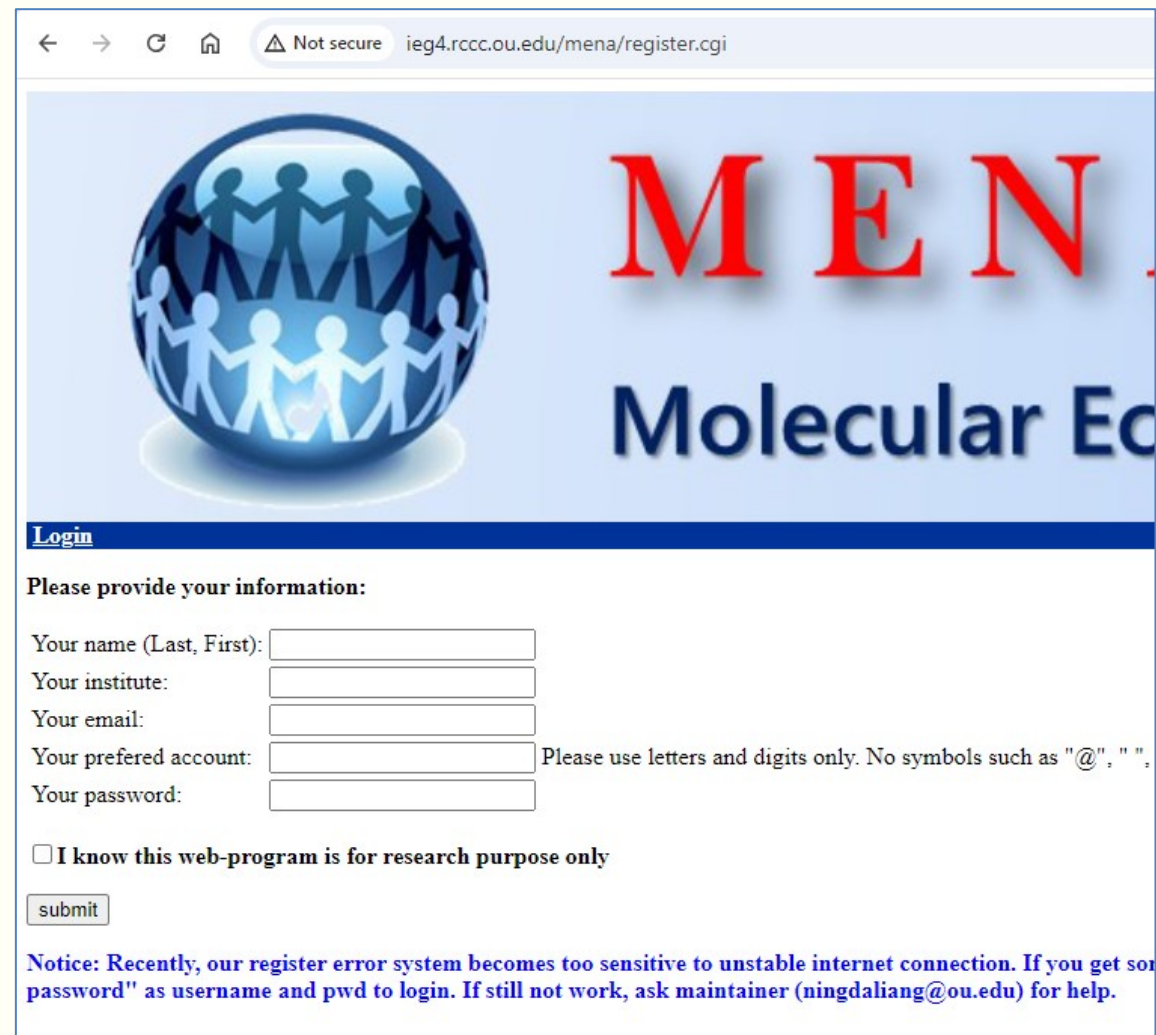
Password:

New user please register [here](#)


Please ensure you visit from new address: <http://ieg4.rccc.ou.edu/mena>

\* Do NOT use any special symbol (e.g. # \$ @) or space in your user name.

\*\* If you forget your password, please contact Daliang (ningdaliang@ou.edu) to reset.



← → ↻ 🏠 Not secure ieg4.rccc.ou.edu/mena/register.cgi



# MENA

## Molecular Ecological

**Login**

Please provide your information:

Your name (Last, First):

Your institute:

Your email:

Your preferred account:  Please use letters and digits only. No symbols such as "@", ".", " ", etc.

Your password:

☐ I know this web-program is for research purpose only

Notice: Recently, our register error system becomes too sensitive to unstable internet connection. If you get some error message like "password" as username and pwd to login. If still not work, ask maintainer (ningdaliang@ou.edu) for help.

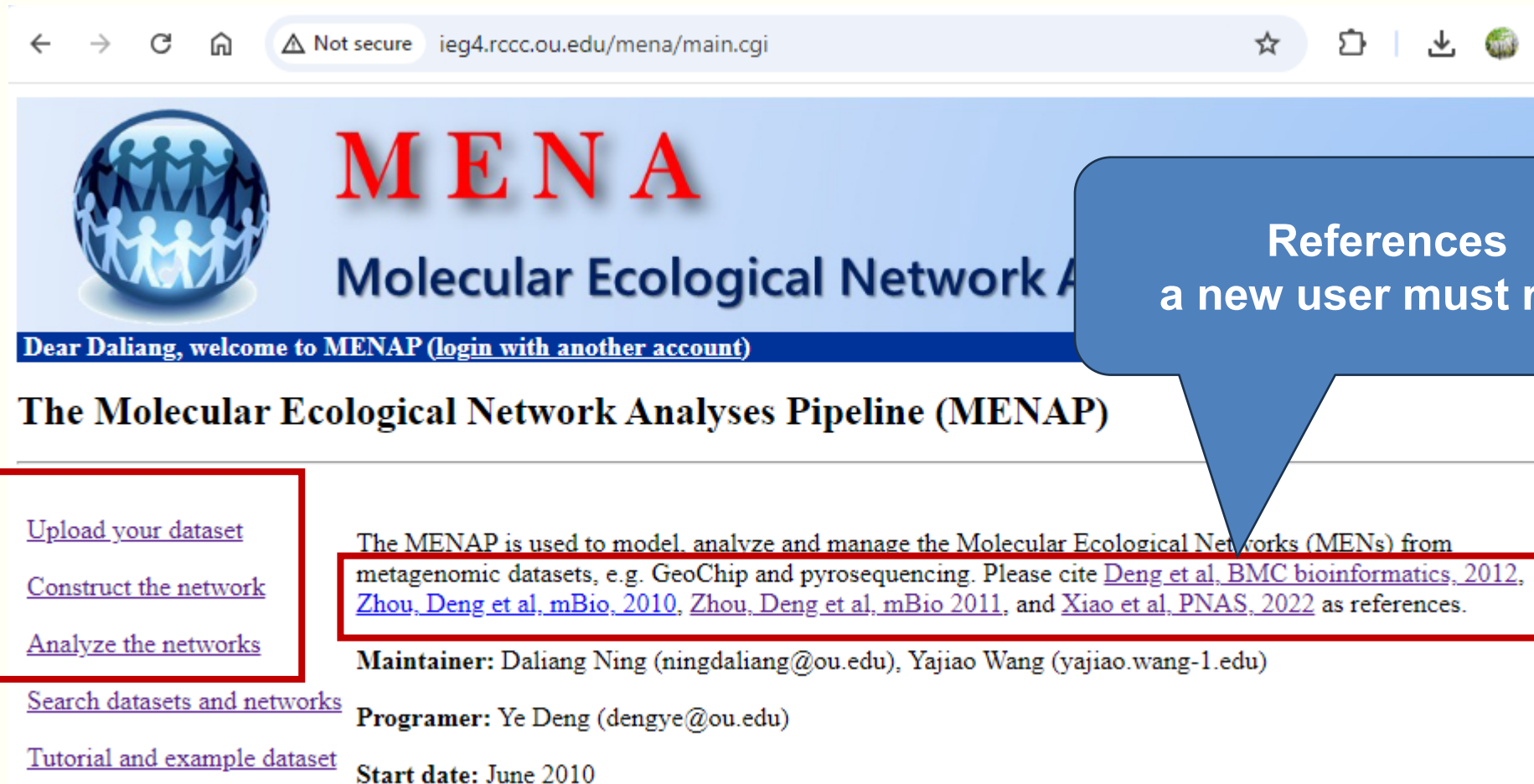
# Registration

- <http://ieg4.rccc.ou.edu/mena/register.cgi>
- ‘Your preferred account’ will be your ‘User Name’
  - Please use letters and digits only. No symbols such as "@", " ", etc. Please do not use your email address.
  - To ensure no conflict with other users, you may need to use a relatively complex account name. Adding time code like 202410061815 can be a good option.
  - Example username: MUS202410061815a
- Unexpected Error message
  - If you get some warning like "this email or username has been used" but you are sure it has not, just go ahead to use what you input to "Your preferred account" and "Your password" as username and password to login <http://ieg4.rccc.ou.edu/mena/>
- For any problem, you can always ask the maintainer for help.




# Log in main page

- <http://ieg4.rccc.ou.edu/mena>
- Once log in, you can see the Three Steps and Recommended References



← → ↻ 🏠 ⚠ Not secure ieg4.rccc.ou.edu/mena/main.cgi ☆ 📁 ⬇ 🌐

 **MENA**  
Molecular Ecological Network Analyses Pipeline

**Dear Daliang, welcome to MENAP (login with another account)**

**The Molecular Ecological Network Analyses Pipeline (MENAP)**

**The Three Steps**

- [Upload your dataset](#)
- [Construct the network](#)
- [Analyze the networks](#)

[Search datasets and networks](#)

[Tutorial and example dataset](#)

**References**  
a new user must read

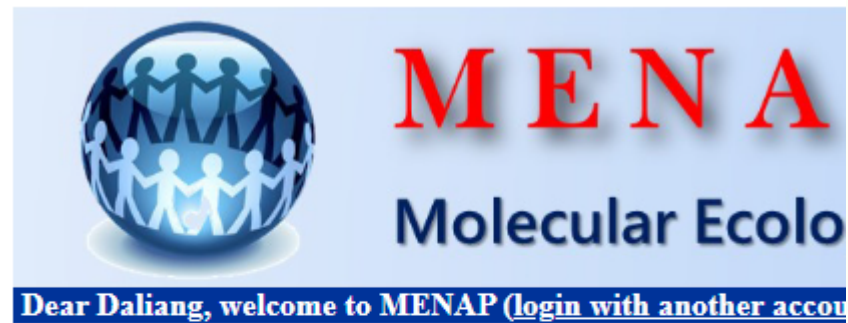
The MENAP is used to model, analyze and manage the Molecular Ecological Networks (MENs) from metagenomic datasets, e.g. GeoChip and pyrosequencing. Please cite [Deng et al, BMC bioinformatics, 2012](#), [Zhou, Deng et al, mBio, 2010](#), [Zhou, Deng et al, mBio 2011](#), and [Xiao et al, PNAS, 2022](#) as references.

**Maintainer:** Daliang Ning (ningdaliang@ou.edu), Yajiao Wang (yajiao.wang-1.edu)

**Programmer:** Ye Deng (dengye@ou.edu)

**Start date:** June 2010

# Upload your dataset (1)



## The Molecular Ecological Network Analysis Platform

[Upload your dataset](#)

[Construct the network](#)

[Analyze the networks](#)

[Search datasets and networks](#)

[Tutorial and example dataset](#)

The MENAP is used to model, analyze and visualize metagenomic datasets, e.g. GeoC, Zhou, Deng et al, mBio, 2010, Zhang et al, mBio, 2010, etc.

**Maintainer:** Daliang Ning (ningdaliang@163.com)

**Programmer:** Ye Deng (dengye@163.com)

**Start date:** June 2010

Main

### Upload your dataset

[Upload your dataset](#)

[Construct the network](#)

[Analyze the networks](#)

[Search datasets and networks](#)

[Tutorial and example dataset](#)

Your data file:

No file chosen

The uploaded file must be **tab-devided text** file and its format should be as same as below (blank data cells are allowed).

ID	Sample1	Sample2	...	SampleN
12345	data11	data12	...	data1N
123456	data21	data22	...	data2N
54321	data31	data32	...	data3N
...				

Please give a descriptive name to your dataset  (need to be complex enough and better include date and initials of your username, e.g. USERA20220414DataA1. Use only letters or letters plus numbers. Do not include any symbols, e.g. #\*,!,:;"')

#### Attention:

- (1) The upload file size could not exceed 30Mb.
- (2) Currently we can not support Mac generated text file. Please switch to Windows or Linux system to generate your text file.
- (3) DO NOT use pure number (e.g. 10) as dataset name.
- (4) Do Not use any blank space or symbols or begin with number in your sample name, taxa name (e.g. OTU ID), or dataset name. Better only letters or letters plus numbers.
- (5) Leave all "zero" (undetectable) cells as blank (empty) cells. Do NOT include any space in the empty cells. Please remove ghost taxa (e.g. OTUs) which are not detected in any sample, unless you have strong rationale to keep them.
- (6) Make the minimum observed value close to but no less than 1. Negative value is not allowed. Do some transformation which would not change correlation coefficient, e.g. multiply the whole matrix by a constant.

#### Reason and Solution for "500 error":

- (1) **Address:** ensure you are using <http://ieg4.rccc.ou.edu/mena/> instead of the old address <http://129.15.40.240/mena/>.
- (2) **Browser:** test an example dataset from [Tutorial and example dataset](#). If it does not work, it is usually browser compatibility issue. Please try **360 browser**, if Firefox, Chrome, or IE does not work. Some users solved the problem by changing to another computer. We feel sorry that Mac can be incompatible now.
- (3) **Format:** if the example dataset works but yours failed, it is usually format issue. Please carefully read the "Attention" above and check your file.


If it still doesn't work, please feel free to contact the maintainers (naijia.xiao@ou.edu). Please include your **username, email, and experiment name** in your email; if you have constructed multiple networks with the same name, please include your network construction time as well.

# Upload your dataset (2)

- OTU or ASV table or Gene (normalized) abundance table
- Tab delimited txt file
- ‘Please give a descriptive name to your dataset’ the name needs to be complex enough to ensure uniqueness
  - e.g., UserA20241006DataC
- Do Not use any blank space or symbols or begin with number in your sample name, taxa name (e.g. OTU ID), or dataset name. Better only letters or letters plus numbers.
  - Good ID examples: OTU1246      SampA453
  - Bad ID examples: 1246   OTU\_1246   453A   453\_A   Samp A 453
- Leave all "zero" (undetectable) cells as blank (empty) cells. Do NOT include any space in the empty cells.
- Please remove ghost taxa (e.g. OTUs) which are not detected in any sample, unless you have strong rationale to keep them.
- Make the minimum observed value close to but no less than 1. Negative value is not allowed.
  - If your data have value  $<1$  or negative values, do some linear transformation which would not change correlation coefficient, e.g., multiply by a constant to bring all observed numbers  $\geq 1$



# Construct the network (1)



**M**  
**Mole**

Dear Daliang, welcome to MENAP ([login with another user](#))

## The Molecular Ecological Network

[Upload your dataset](#)

[Construct the network](#)

[Analyze the networks](#)

[Search datasets and networks](#)

[Tutorial and example dataset](#)

The MENAP pyrosequencing dataset 2022 as reference

**Maintainer:**

**Programmer:**

**Start date:**

Main

### Select Dataset

search by:

order by:

Please select a dataset to construct Molecular Ecological Network (MEN)

	User	Experiment name	Upload time	File name	Number of samples	Number of genes
<input checked="" type="radio"/>	Daliang	warmtest240922A	2024-09-22 18:49:31	warming-34.txt	14	976
<input type="radio"/>	Daliang	M120d20240723a	2024-07-23 18:13:58	M12p0.txt	8	550

# Construct the network (2)

Main > Select dataset

## Settings of MEN construction

Data Preparation

Settings of RMT  
(Random Matrix Theory)

**Majority:** Only keep the genes/OTUs with  in total 14 samples

**Missing data:**

- ☐ Keep blank
- ☐ Fill with
- ☒ Only fill with  in blanks with paired valid values
- ☐ Fill with ten Nearest Neighbors (Troyanskaya et al. 2001, Bioinformatics)

**Transformation:**

- ☒ Centered log-ratio transform (CLR, see a summary by Dr. Tolosana-Delgado [here](#), recommended for [compositional data](#)).
- ☐ Logarithm transformation (recommended for non-compositional data)
- ☐ No transformation
- ☒ Pearson Correlation Coefficient (recommended)
- ☐ Mutual Information
- ☐ Spearman's Rho using  $r_s = 1 - \frac{6 \sum d_i^2}{n(n^2 - 1)}$  (deprecated) where  $rg_X, rg_Y$  is the rank of the raw score,  $d_i = rg(X_i) - rg(Y_i)$  is the difference, and  $n$  is the number of observations

**Similarity matrix:**

- ☐ Spearman's Rho using  $r_s = \rho_{rg_X, rg_Y} = \frac{\text{cov}(rg_X, rg_Y)}{\sigma_{rg_X} \sigma_{rg_Y}}$
- ☐ Pearson correlation in time-series (allow  time point lagging) (samples with biological replicates which must upload a sample description file [click here](#))
- ☐ Pearson correlation in time-series (allow  time point lagging) (the samples already followed time-series and without biological replicates)

**Calculation order:**

- ☒ Decrease the cutoff from top
- ☐ Increase the cutoff from
- ☐ From  to  with step  (can't loop more than 50 times)

**Scan speed:**

- ☒ Regress Poisson distribution only(quiicker)
- ☐ Regress GOE and Poisson (slower)

You can change this number. Recommend to use 50% - 80% of the total sample number, or a number you can well justify.

Recommend to use this option. You may change the 'fill-with' number, but it must be lower than the minimum observed number, but not so low that it can lead to false correlation. Default is 0.01, assuming the minimum observed number is 1.

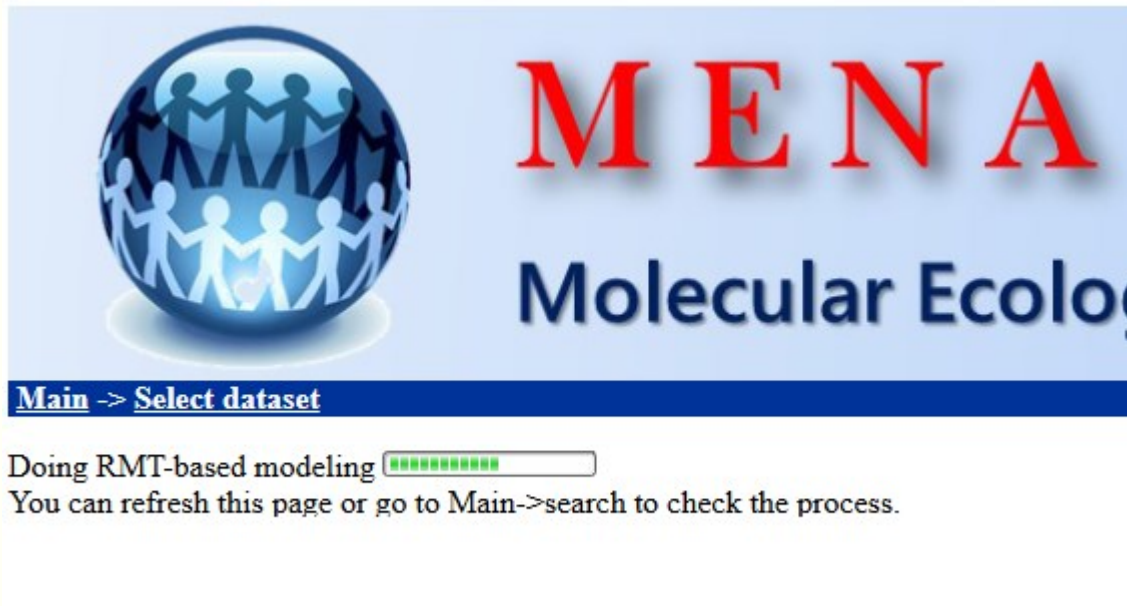
This is recommended, to mitigate the 'compositional data' effect on correlation estimation. If your data is not compositional, e.g., absolute abundance data, try other options.

**DO NOT use this Spearman option. Use the next one if you want to use Spearman.**

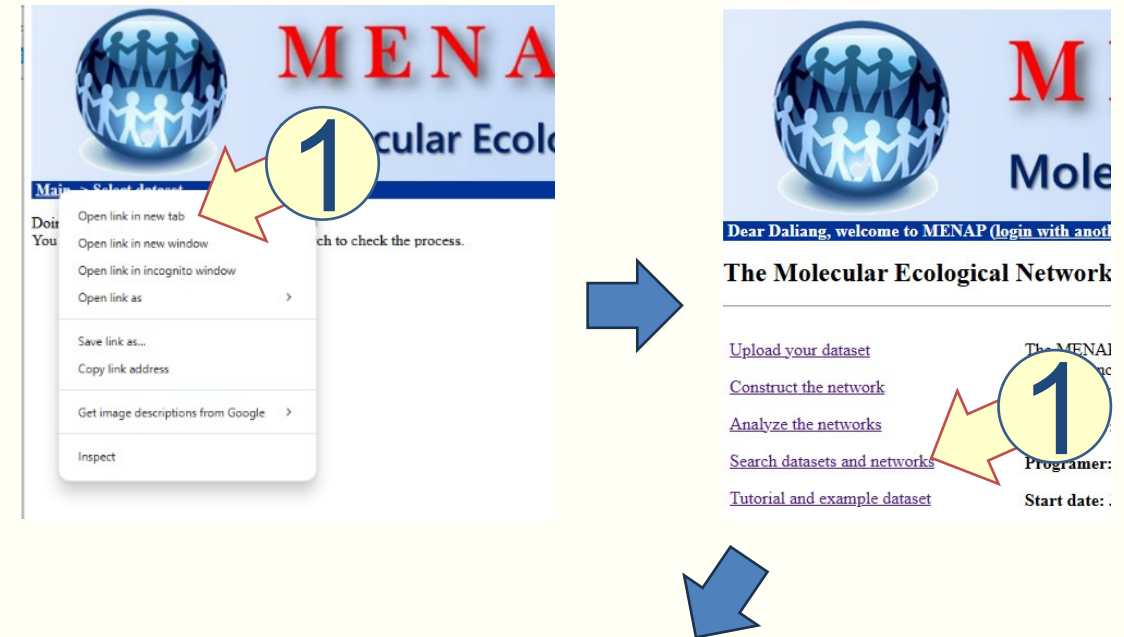
submit

# Construct the network (3)

When the page does not show 'You can refresh this page ...', DO NOT close or refresh the page, but wait.



When waiting, you can right-click 'Main' → 'open link in new Tab', then in a new Tab, click 'Search datasets and networks' to check progress



Once the 'Status' shows 'done', you may refresh the original page, or click on the 'Name' of the dataset.

[Main](#)

**Datasets Summary**

[Click here, go to networks summary](#)

search by:  date  submit

No	Name	Time	#Samples	#Genes	Settings	Status
1	<a href="#">warmtest20241123a</a>	2024-11-23 10:45:21	14	228	majority=7; missing_fill=fill_paired(0.0100); logarithm=c; similarity=pcc;	done <input type="checkbox"/>
2	<a href="#">warmtest240922A</a>	2024-09-22 18:49:48	14	228	majority=7; missing_fill=fill_paired(0.0100); logarithm=c; similarity=pcc;	done <input type="checkbox"/>

# Construct the network (4)

[Main](#) -> [Select dataset](#)

RMT-based modeling has been done on "warmingtest20241123a" dataset.

Please choose a cutoff according to the results below for similarity matrix to construct the network.

Cutoff	Chi-square test on Poisson distribution	p value
0.890	167.978	0.000
0.880	70.216	0.172
0.870	53.215	0.720
0.860	72.251	0.133
0.850	65.037	0.306
0.840	67.063	0.248
0.830	66.426	0.265
0.820	109.416	0.000
0.810	69.285	0.193
0.800	63.660	0.349
0.790	63.187	0.364
0.780	74.178	0.103
0.770	81.031	0.037
0.760	86.087	0.015
0.750	95.784	0.002
0.740	124.967	0.000

0.030	123.438	0.000	p>0.001
0.020	127.558	0.000	p>0.01
0.010	118.413	0.000	p>0.05

GBD-based RMT cutoffs for "warmingtest20241123a Pearson Correlation.txt": upper=0.8,best=0.77,lower=0.75

☒ Use [iDIRECT](#) to remove indirect relationships? Hover for more

The cutoff:

Raw similarity matrix: [download](#) (Only the upper-right triangle of the full matrix. It contains no OTU ID.)

Condensed OTU table: [download](#) (OTU ID in the same order as the correlation matrix)

Custom similarity matrix: [upload](#)

'RMT cutoffs based on GBD function' is more recommended now. Theoretically, any number between the 'upper' and 'lower' values can be used as a good cutoff.

- If you only construct one network for your project, please use the 'best' value;
- If you will compare several different networks, you better get this range ('upper' to 'lower') for each network first, then use a cutoff value which can fall into this range for every network. Then, by using the same cutoff value, those networks can be more comparable.

'iDIRECT' is recommended to remove the 'indirect' associations, to make the result network only include 'direct' associations.

- If you selected 'iDIRECT' once but want to try not to use 'iDIRECT', you need to upload your file again with another name.



# Construct the network (5)

[Main](#) -> [Select dataset](#)

RMT-based modeling has been done on "warmingtest20241123a" dataset.

Please choose a cutoff according to the results below for similarity matrix to construct the network.

Cutoff	Chi-square test on Poisson distribution	p value
0.890	167.978	0.000
0.880	70.216	0.172
0.870	53.215	0.720
0.860	72.251	0.133
0.850	65.037	0.306
0.840	67.063	0.248
0.830	66.426	0.265
0.820	109.416	0.000
0.810	69.285	0.193
0.800	63.660	0.349
0.790	63.187	0.364
0.780	74.178	0.103
0.770	81.031	0.037
0.760	86.087	0.015
0.750	95.784	0.002
0.740	124.967	0.000

0.030	123.438	0.000	p>0.001
0.020	127.558	0.000	p>0.01
0.010	118.413	0.000	p>0.05

This is from our old algorithm, **NOT recommended any more**, which did not work for a few datasets.

Here, the default value is from our old algorithm, **NOT recommended any more**. Please consider using the GBD-based RMT cutoffs as described on previous page.

GBD-based RMT cutoffs for "warmingtest20241123a Pearson Correlation.txt": upper=0.8,best=0.77,lower=0.75

☒ Use [iDIRECT](#) to remove indirect relationships? Hover for more

The cutoff:

Raw similarity matrix: [download](#) (Only the upper-right triangle of the full matrix. It contains no OTU ID.)

Condensed OTU table: [download](#) (OTU ID in the same order as the correlation matrix)

Custom similarity matrix: [upload](#)

# Construct the network (6)

[Main](#) -> [Select dataset](#)

RMT-based modeling has been done on "warmingtest20241123a" dataset.

Please choose a cutoff according to the results below for similarity matrix to construct the network.

Cutoff	Chi-square test on Poisson distribution	p value
0.890	167.978	0.000
0.880	70.216	0.172
0.870	53.215	0.720
0.860	72.251	0.133
0.850	65.037	0.306
0.840	67.063	0.248
0.830	66.426	0.265
0.820	109.416	0.000
0.810	69.285	0.193
0.800	63.660	0.349
0.790	63.187	0.364
0.780	74.178	0.103
0.770	81.031	0.037
0.760	86.087	0.015
0.750	95.784	0.002
0.740	124.967	0.000

0.030	123.438	0.000	<span style="background-color: #ffcccc;">p&gt;0.001</span>
0.020	127.558	0.000	<span style="background-color: #ffcccc;">p&gt;0.01</span>
0.010	118.413	0.000	<span style="background-color: #ffcccc;">p&gt;0.05</span>

GBD-based RMT cutoffs for "warmingtest20241123a Pearson Correlation.txt": upper=0.8,best=0.77,lower=0.75

☒ Use [iDIRECT](#) to remove indirect relationships? Hover for more

The cutoff:

Raw similarity matrix: [download](#) (Only the upper-right triangle of the full matrix. It contains no OTU ID.)

Condensed OTU table: [download](#) (OTU ID in the same order as the correlation matrix)

Custom similarity matrix: [upload](#)

Raw similarity matrix: the upper-right triangle of the 'association matrix' before applying any cutoff, where the taxa IDs are not included but in the exact order as the taxa IDs in the 'condensed OTU table'.

Condensed OTU table: the community composition matrix, each row is a taxon, each column is a sample, after applying the 'majority' filter and data transformation.

This is where you may upload a similarity matrix (association matrix) which results from your own algorithm that is not available in MENAP.

- Remember to follow the format requirement.

# Construct the network (7)

Revise the cutoff according to GBD-based RMT cutoffs. See notes in the previous slide .

GBD-based RMT cutoffs for "warmingtest20241123a Pearson Correlation.txt": upper=0.8,best=0.77,lower=0.75

☒ Use [iDIRECT](#) to remove indirect relationships? Hover for more

The cutoff:

Raw similarity matrix: [download](#) (Only the upper-right triangle of the full matrix. It contains no OTU ID.)

Condensed OTU table: [download](#) (OTU ID in the same order as the correlation matrix)

Custom clarity matrix: [upload](#)




**Main**

The number of nodes: 120  
 The number of links: 163  
 The average path: 4.558  
 R square of power-law: 0.924

A rough summary will show, it is good to go to the Main page and start next step.

# Download matrixes (1)

- Download correlation/similarity matrix and condensed OTU table
  - First, 'Search datasets and networks' -> select the dataset



**M**  
**Mole**

**Dear Daliang, welcome to MENAP (login with another user)**

**The Molecular Ecological Network**

[Upload your dataset](#)

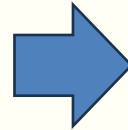
[Construct the network](#)

[Analyze the networks](#)

[Search datasets and networks](#)

[Tutorial and example dataset](#)

The MENAP  
pyrosequencing  
2022 as reference  
Maintainer:  
Programmer:  
Start date:



**Main**

**Datasets Summary**

[Click here go to networks summary](#)

search by:  date

No	Name	#Samples	#Genes	Settings	Status
1	<a href="#">warmingtest20241123a</a>	14	228	majority=7; missing_fill=fill_paired(0.0100); logarithm=c; similarity=pcc;	done <input type="checkbox"/>
2	<a href="#">warmtest240922A</a>	14	228	majority=7; missing_fill=fill_paired(0.0100); logarithm=c; similarity=pcc;	done <input type="checkbox"/>



# Download matrixes (2)

## • Download the similarity matrix and condensed OTU table

### ▪ 'Condensed OTU table'

- the OTU/ASV/gene table after applying 'majority' rule, i.e., without taxa detected in too few samples.

### ▪ 'Raw similarity matrix'

- the pairwise correlation coefficients, but those with absolute values lower than the default cutoff are set to zero. If you need all the values, contact the administrator and provide your user name and the dataset name.
- The file has only upper-right triangle values of the full matrix and no ID (but the rows are in the same order as the condensed OTU table)
  - An example R code to transform the data file to a full matrix  
<http://ieg4.rccc.ou.edu/MENA/download/MENA.similarity.matrix.convert.r.txt>

GBD-based RMT cutoffs for "warmingtest20241123a Pearson Correlation.txt": upper=0.8,best=0.77,lower=0.75

☒ Use [iDIRECT](#) to remove indirect relationships? Hover for more

The cutoff:

Raw similarity matrix: [download](#) (Only the upper-right triangle of the full matrix. It contains no OTU ID.)


Condensed OTU table: [download](#) (OTU ID in the same order as the correlation matrix)

Custom similarity matrix: [upload](#)

Right-click '*download*' and 'Save link as ...' to download the tables.

# Analyze the networks

Please run the tools **one by one from top to bottom**, except the 'randomize the network ...' which is time consuming and can be run at the last.



**M**  
**Mole**

Dear Daliang, welcome to MENAP (login with another user)

## The Molecular Ecological Network

[Upload your dataset](#)

[Construct the network](#)

[Analyze the networks](#)

[Search datasets and networks](#)

[Tutorial and example dataset](#)

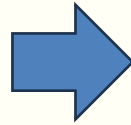
The MENAP pyrosequencing data from 2022 as reference

Maintainer: ...

Programmer: ...

Start date: ...

**1**



**Main**

### Network analyses

[Upload your dataset](#)

[Construct the network](#)




[Analyze the networks](#)

[Search datasets and networks](#)



[Tutorial and example dataset](#)

You had better run the following analyses one by one from top to bottom. But you may do "Randomize the network structure" at last, as it can be time consuming.





#### Network reports

-  Global Network properties
-  Individual nodes' centrality
-  Module Separation and modularity calculation

#### Network visualization

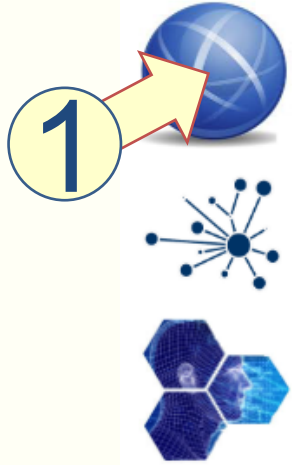
-  Simple network plot (un-editable)
-  Output for the [Cytoscape](#) software visualization

#### Network statistics

-  Fit three power-law models (regular power law, exponential law and truncated power law)
-  Randomize the network structure and then calculate network properties (Updated: Modularity values from random networks have been added.)
-  Calculate Gene/OTU significances (GS) with environmental traits and then use Mantel test to check the correlations between GS and network connectivity
-  Module-EigenGene analyses

# Global Network Properties

## Network reports



Global Network properties

Individual nodes' centrality

Module Separation and modularity calculation



Main -> Analysis

Select Network to do global\_property

search by:  date   
 order by:  date  descending  submit

Please select a Molecular Ecological Network (MEN)

	User	Experiment name	Construction time	RMT threshold	# nodes	# links
<input checked="" type="checkbox"/>	Daliang	warmingtest20241123a	2024-11-23 11:29:18	0.7700	120	163
<input type="checkbox"/>	Daliang	M120d20240723a	2024-07-23 18:26:40	0.9000	136	110

Submit



If the dataset is very big, you may need to wait for some hours.

Main -> Analysis

## Network global properties

This is your first time to run parameter calculations on warmingtest20241123a at 0.77 threshold. It will take a few minutes...

Network Indexes	warmingtest20241123a(0.77)
Total nodes	120
Total links	163
R square of power-law	0.924
Average degree (avgK)	2.717
Average clustering coefficient (avgCC)	0.001
Average path distance (GD)	4.558
Geodesic efficiency (E)	0.278
Harmonic geodesic distance (HD)	3.602
Maximal degree	19
Nodes with max degree	OTU1094
Centralization of degree (CD)	0.139
Maximal betweenness	1485.297
Nodes with max betweenness	OTU1094
Centralization of betweenness (CB)	0.193
Maximal stress centrality	11108
Nodes with max stress centrality	OTU1094
Centralization of stress centrality (CS)	1.434
Maximal eigenvector centrality	0.413
Nodes with max eigenvector centrality	OTU1094
Centralization of eigenvector centrality (CE)	0.369
Density (D)	0.023
Reciprocity	1
Transitivity (Trans)	0.004
Connectedness (Con)	0.655
Efficiency	0.977
Hierarchy	0
Lubness	1

Copy paste the table to a spreadsheet

# Individual Nodes Centrality (node attributes)

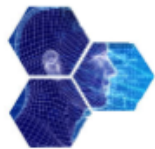
## Network reports



Global Network properties



Individual nodes' centrality



Module Separation and modularity calculation



## Main -> Analysis

### Select Network to do individual\_property

search by:

order by:

Please select a Molecular Ecological Network (MEN)

	User	Experiment name	Construction time	RMT threshold	# nodes	# links
<input checked="" type="radio"/>	Daliang	warmingtest20241123a	2024-11-23 11:29:18	0.7700	120	163
<input type="radio"/>	Daliang	M120d20240723a	2024-07-23 18:26:40	0.9000	136	110

## Main -> Analysis

### Network nodes' centrality indexes

Node Name	node.degree	node.betw	node.stress	node.evcent	Clustering.Coefficient
OTU1	10	641.712	6278	0.266	0
OTU1006	8	373.113	4306	0.204	0
OTU1033	1	0	0	0	0
OTU1035	1	0	0	0.023	0
OTU1060	3	33.451	214	0.028	0
OTU1068	2	2	2	0	0
OTU107	1	0	0	0.005	0
OTU1076	2	59.287	668	0.051	0
OTU1088	5	376.834	2956	0.002	0
OTU1094	19	1485.297	11108	0.412	0.005
OTU783	1	0	0	0.000	0
OTU896	4	23.081	258	0.096	0
OTU898	2	0.84	7	0.070	0
OTU96	1	0	0	1.324e-05	0

You can download this table [here](#), and then open it in Excel.

- Open link in new tab
- Open link in new window
- Open link in incognito window
- Open link as >
- Save link as...
- Copy link address
- Get image descriptions from Google >
- Inspect

Right-click 'here' and 'Save link as ...' to download the table.



# Module Separation and Modularity

## Network reports



Global Network properties



Individual nodes' centrality



Module Separation and modularity calculation

1



Main -> Analysis

## Select Network to do modularity

search by:

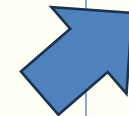
order by:

Please select a Molecular Ecological Network (MEN)

	User	Experiment name	Construction time	RMT threshold	# nodes	# links
<input checked="" type="radio"/>	Daliang	warmingtest20241123a	2024-11-23 11:29:18	0.7700	120	163
<input type="radio"/>	Daliang	M120d20240723a	2024-07-23 18:26:40	0.9000	136	110

Submit

3



Main -> Analysis

## Settings of module separation

Summary of your data:

Experiment name	warmingtest20241123a
Threshold	0.77
# nodes	120
# links	163

4

Separation method:

☒ greedy modularity optimization

☐ short random walks

☐ leading eigenvector of the community matrix

Other options:

☒ Calculate Z and P values for all nodes

☐ Calculate ZP values and plot them into a scatter plot



Main -> Analysis

## Network module separation and modularity calculation

This is your first time to run parameter calculations on warmingtest20241123a at 0.77 threshold. It will take a few minutes...

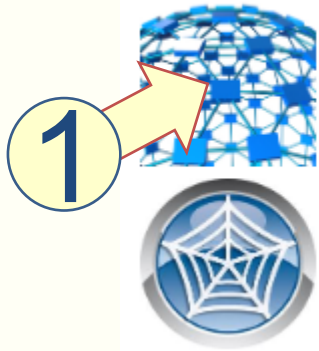
#module: 14				
modularity: 0.615				
No	ID	No. module	Zi	Pi
1	OTU1	2	1.224	0.54
2	OTU1006	3	0.079	0.718
120		1	-0.691	0

You can download the [table](#), and then open it in Excel.

Right-click 'here' and 'Save link as ...' to download the table.

# Simple network plot (un-editable)

## Network visualization



Simple network plot (un-editable)

Output for the [Cytoscape](#) software visualization



## Main -> Analysis

### Select Network to do Simple\_plot

search by:  date

order by:  date  descending  submit

2 Please select a Molecular Ecological Network (MEN)

	User	Experiment name	Construction time	RMT threshold	# nodes	# links
<input checked="" type="radio"/>	Daliang	warmingtest20241123a	2024-11-23 11:29:18	0.7700	120	163
<input type="radio"/>	Daliang	M120d20240723a	2024-07-23 18:26:40	0.9000	136	110

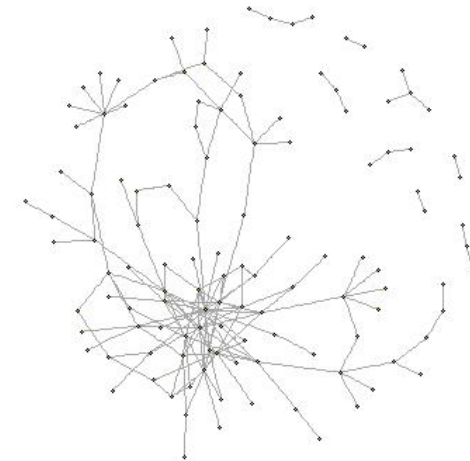
Submit

3

## Main -> Analysis

### The simple plot of network

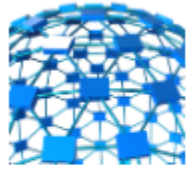
The figure:



This is a quick overview of the network.

# Output for the Cytoscape software visualization

## Network visualization



Simple network plot (un-editable)



Output for the [Cytoscape](#) software visualization



[Main -> Analysis](#)

## Select Network to do Cytoscape\_preparation

search by:  date

order by:  date  descending  submit

2 Please select a Molecular Ecological Network (MEN)

	User	Experiment name	Construction time	RMT threshold	# nodes	# links
<input checked="" type="radio"/>	Daliang	warmingtest20241123a	2024-11-23 11:29:18	0.7700	120	163
<input type="radio"/>	Daliang	M120d20240723a	2024-07-23 18:26:40	0.9000	136	110

Submit

3

[Main -> Analysis](#)

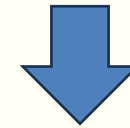
## Output the files for the Cytoscape visualization

4 You have already ran these module-separation methods. Please choose one:

☒ greedy modularity optimization

submit

5



[Main -> Analysis](#)

## Output the files for the Cytoscape visualization

The network file [download here](#)

The node attribute file [download here](#)

The edge attribute file [download here](#)

\*The Cytoscape software can be downloaded [here](#). Its online tutorial is [here](#).

Right-click each file link and 'Save link as ...' to download the table.

# Use Cytoscape to visualize your network

- Download and install Cytoscape
  - <https://cytoscape.org/>
- Check the online tutorial from Cytoscape
  - <https://github.com/cytoscape/cytoscape-tutorials/wiki>
- Below is a simple example
  - Using Cytoscape 3.10.3



# Use Cytoscape: prepare your files

- Download files from MENAP function 'Output for the Cytoscape software visualization'

**Main -> Analysis**

**Output the files for the Cytoscape visualization**

The network file [download here](#)

The node attribute file [download here](#)

The edge attribute file [download here](#)

\*The Cytoscape software can be downloaded [here](#). Its online tutorial is [here](#).



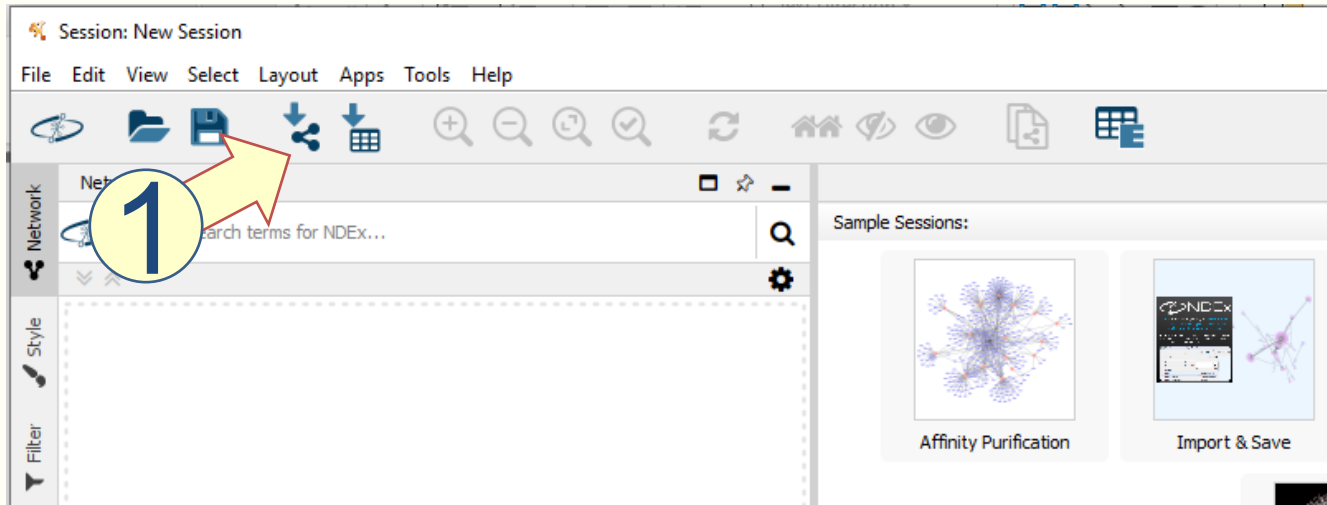
Name	Date modified	Type	Size
warmingtest20241123a 0.77 edge_attribute.txt	11/23/2024 12:48 PM	Text Document	5 KB
warmingtest20241123a 0.77 node_attribute.txt	11/23/2024 12:48 PM	Text Document	8 KB
warmingtest20241123a 0.77.sif	11/23/2024 12:48 PM	SIF File	4 KB

- Use Excel to edit the '...node\_attribute.txt' or '...edge\_attribute.txt' if you want to add more attributes to show in the network figure.
  - For example, add the phylum information

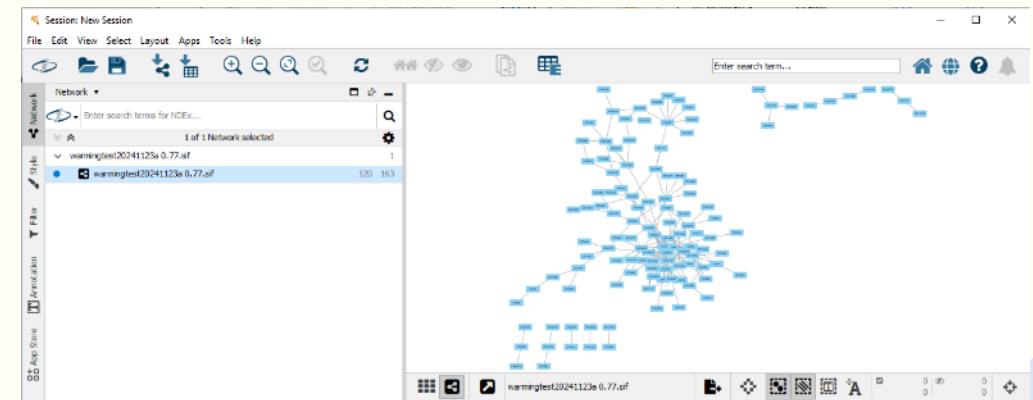
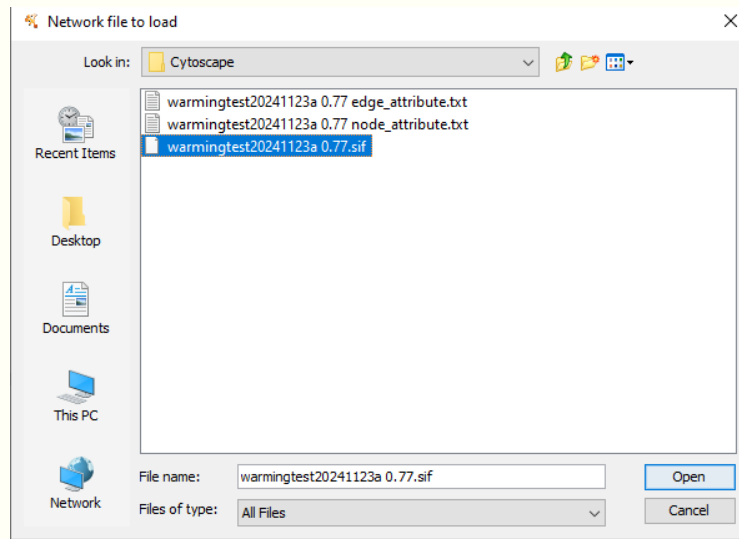
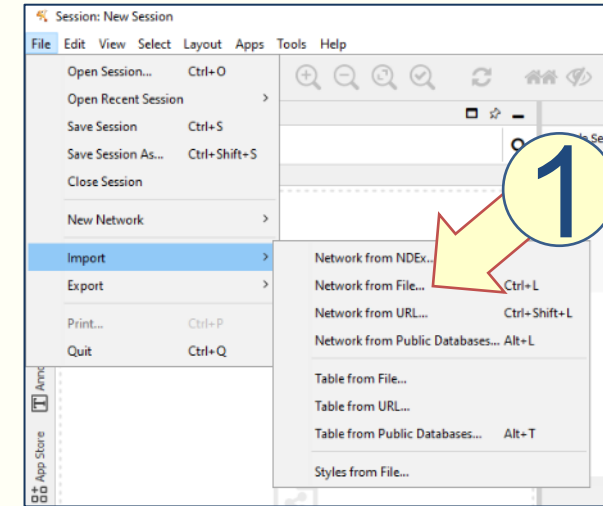
	A	B	C	D	E	F	G	H	I	J	K
1	Name	node.degr	node.betw	node.stres	node.evce	Clustering	No. modu	Zi	Pi	Phylum	
2	OTU1	10	641.7128	6278	0.266659	0	2	1.224745	0.54	Firmicutes	
3	OTU1006	8	373.1139	4306	0.204756	0	3	0.079057	0.71875	Bacteroidetes	
4	OTU1033	1	0	0	0	0	12	0	0	Actinobacteria	
5	OTU1035	1	0	0	0.023869	0	0	-0.87706	0	Proteobacteria	
6	OTU1060	3	33.45186	214	0.028578	0	0	0	0.444444	Verrucomicrobia	
7	OTU1068	2	2	2	0	0	6	1	0	Fusobacteria	
8	OTU107	1	0	0	0.005383	0	1	-0.69109	0	Cyanobacteria	
9	OTU1076	2	59.28701	668	0.051311	0	1	-0.69109	0.5	Firmicutes	
10	OTU1088	5	376.8348	2956	0.002985	0	4	2.183063	0	Firmicutes	
11	OTU1094	19	1485.297	11108	0.412795	0.005848	2	3.265986	0.592798	Firmicutes	

# Use Cytoscape: import data files (1)

- Import the network file '...sif'

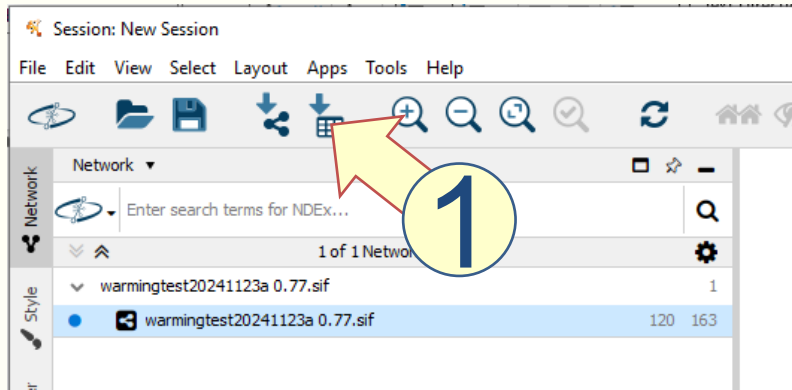


OR

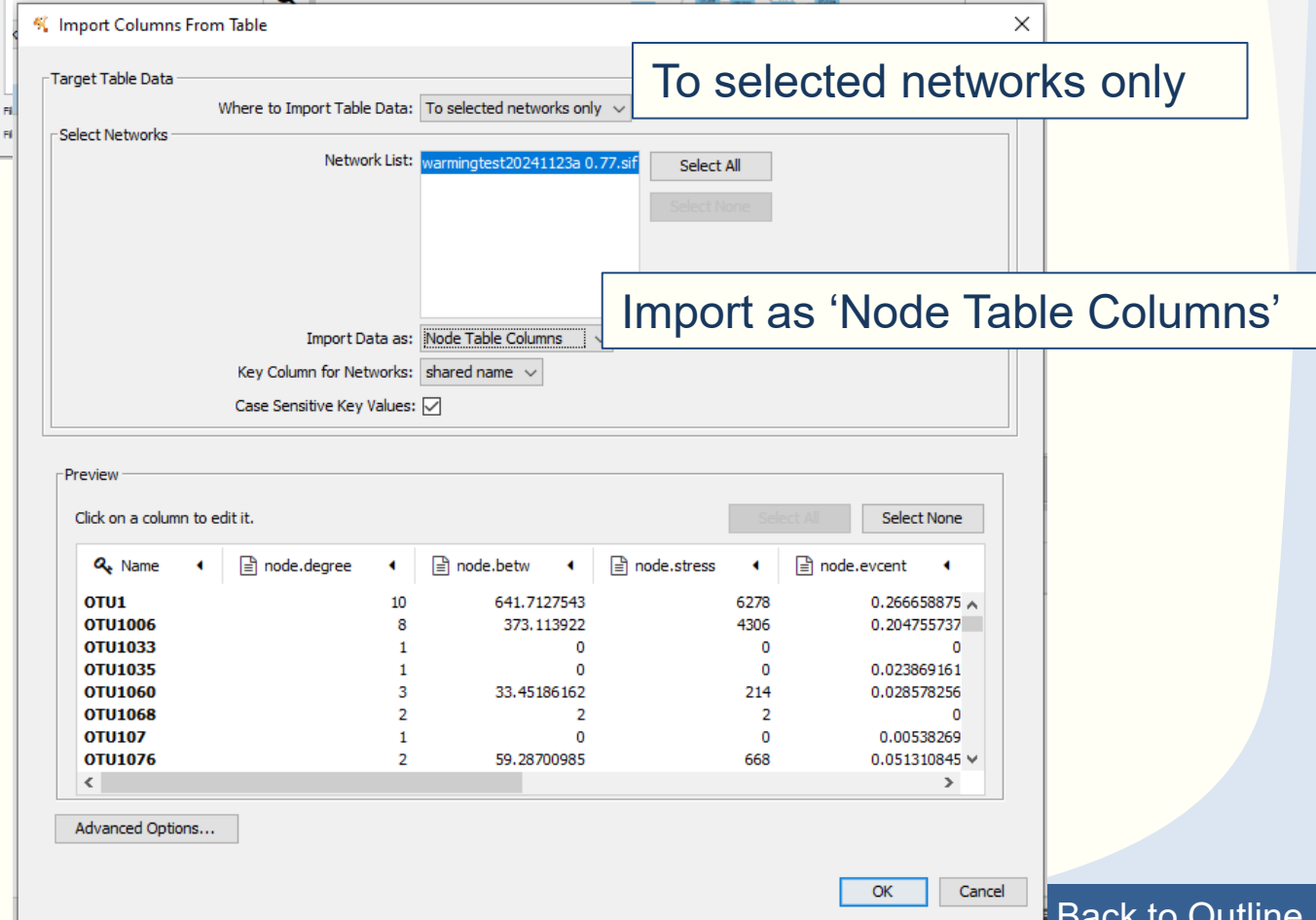
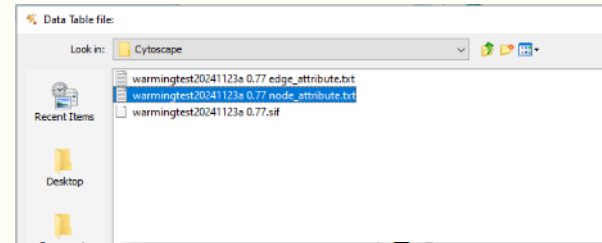
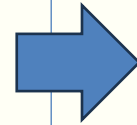
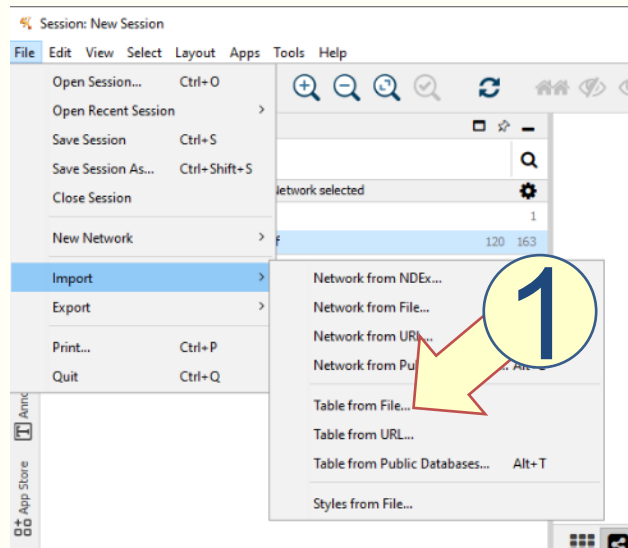


# Use Cytoscape: import data files (2)

- Import the '...node\_attribute.txt' file

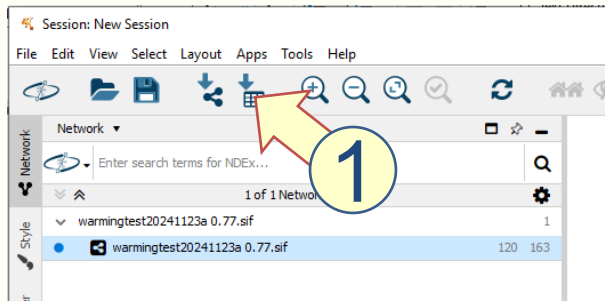


OR

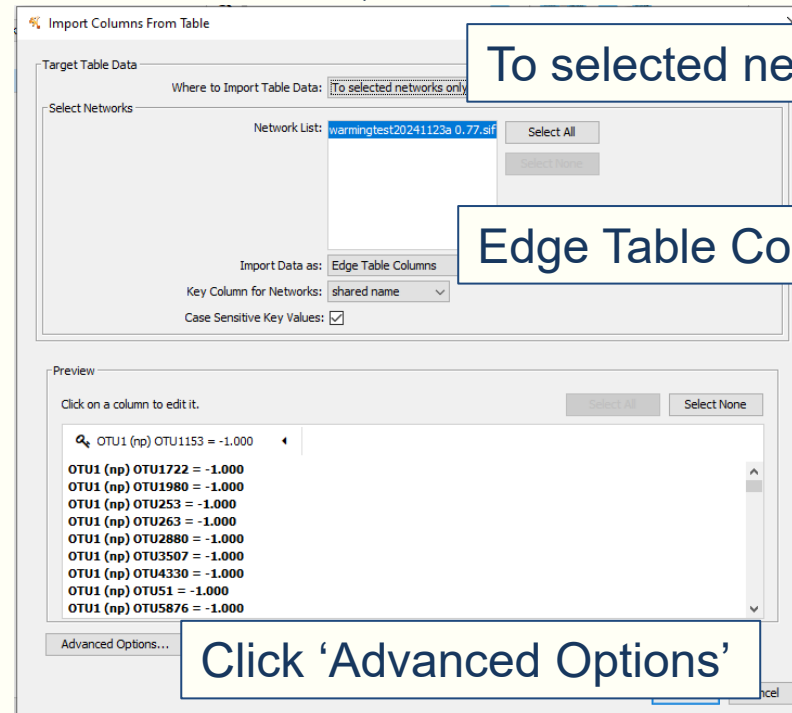
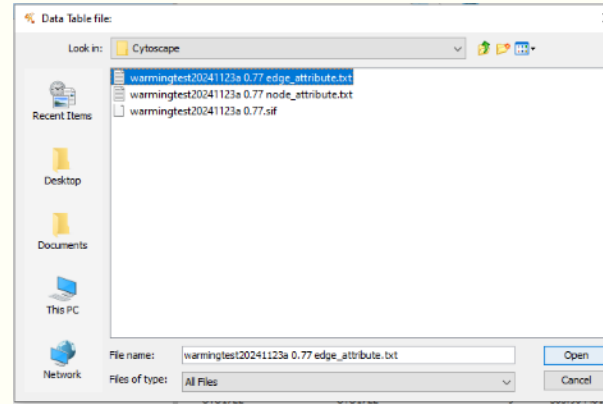
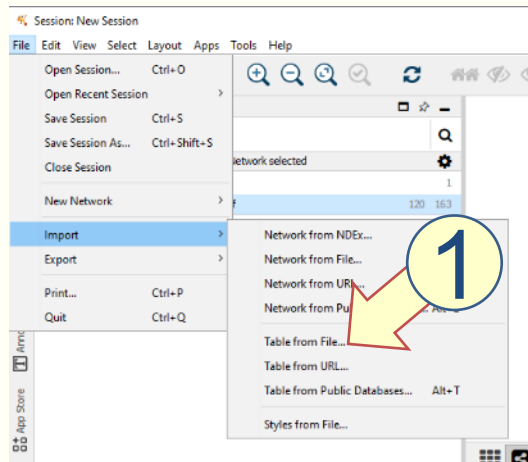


# Use Cytoscape: import data files (3)

- Import the '...edge\_attribute.txt' file



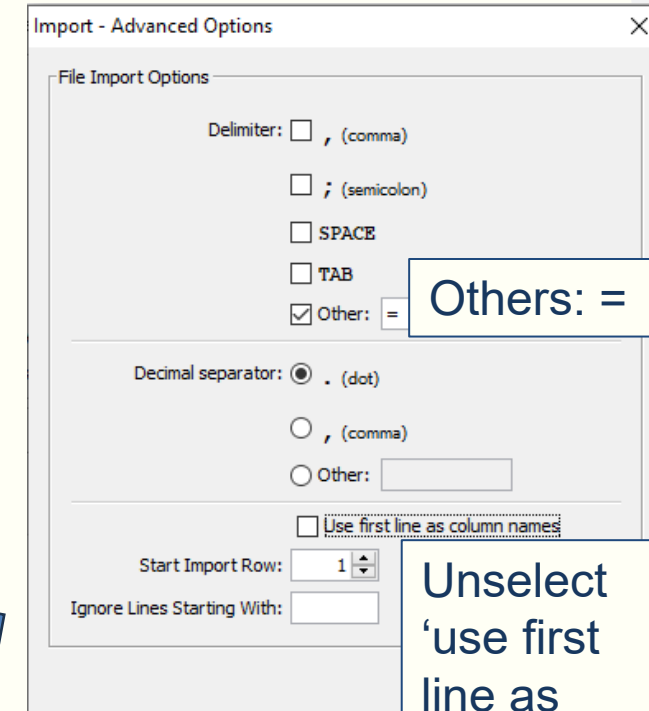
OR



To selected networks only

Edge Table Columns

Click 'Advanced Options'



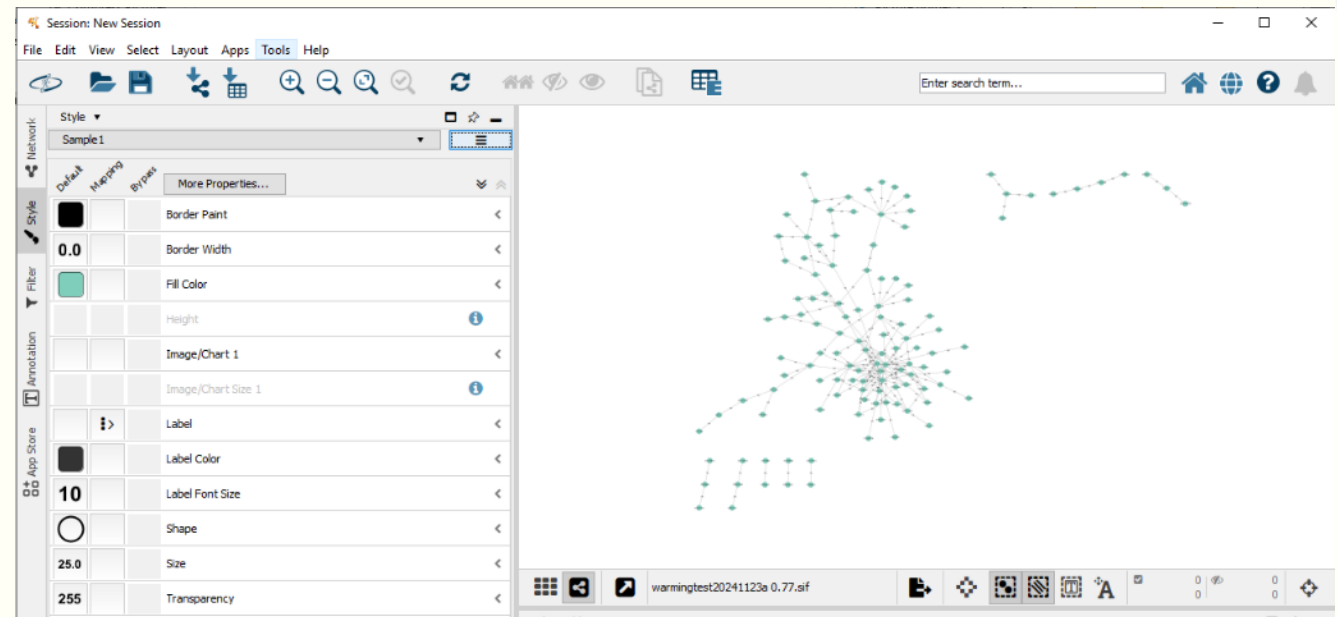
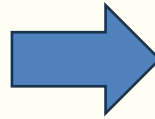
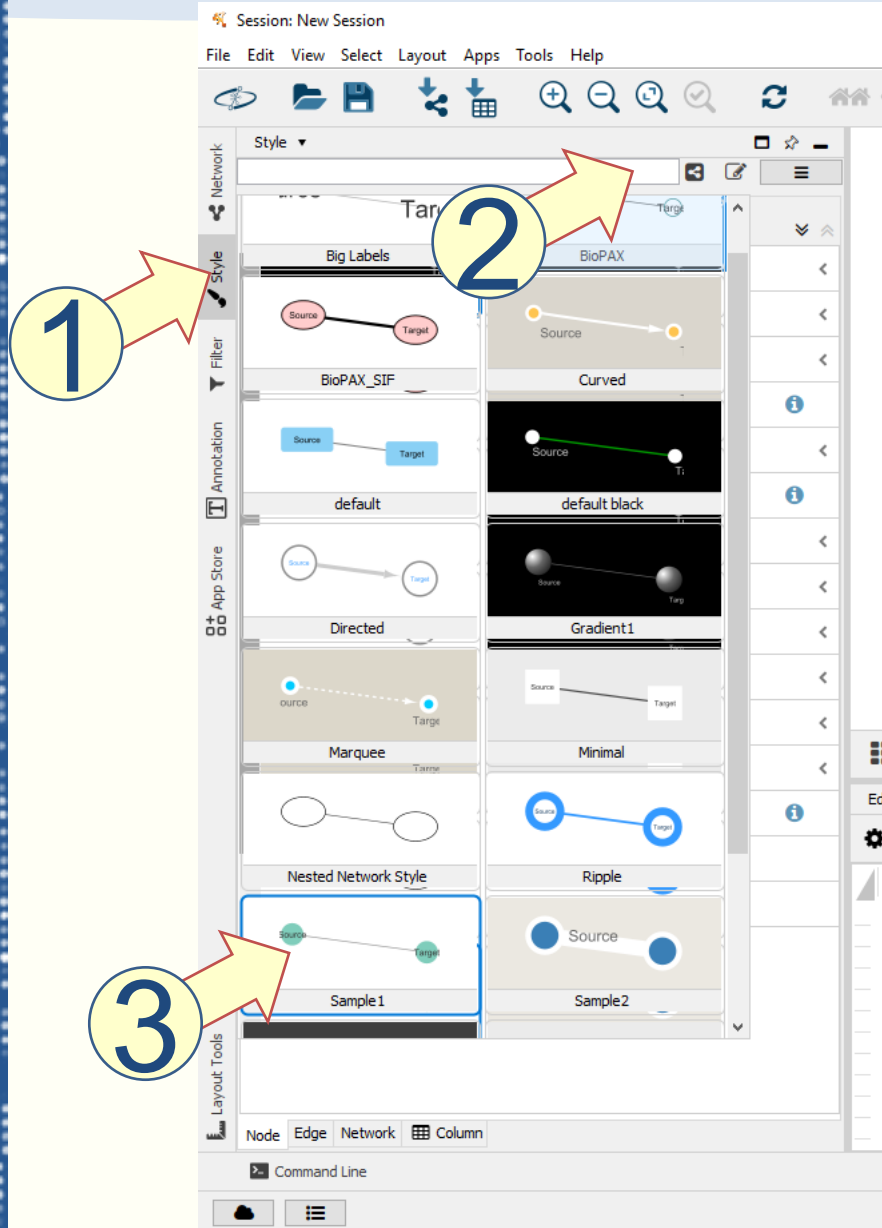
Others: =

Unselect  
'use first  
line as  
column  
names'



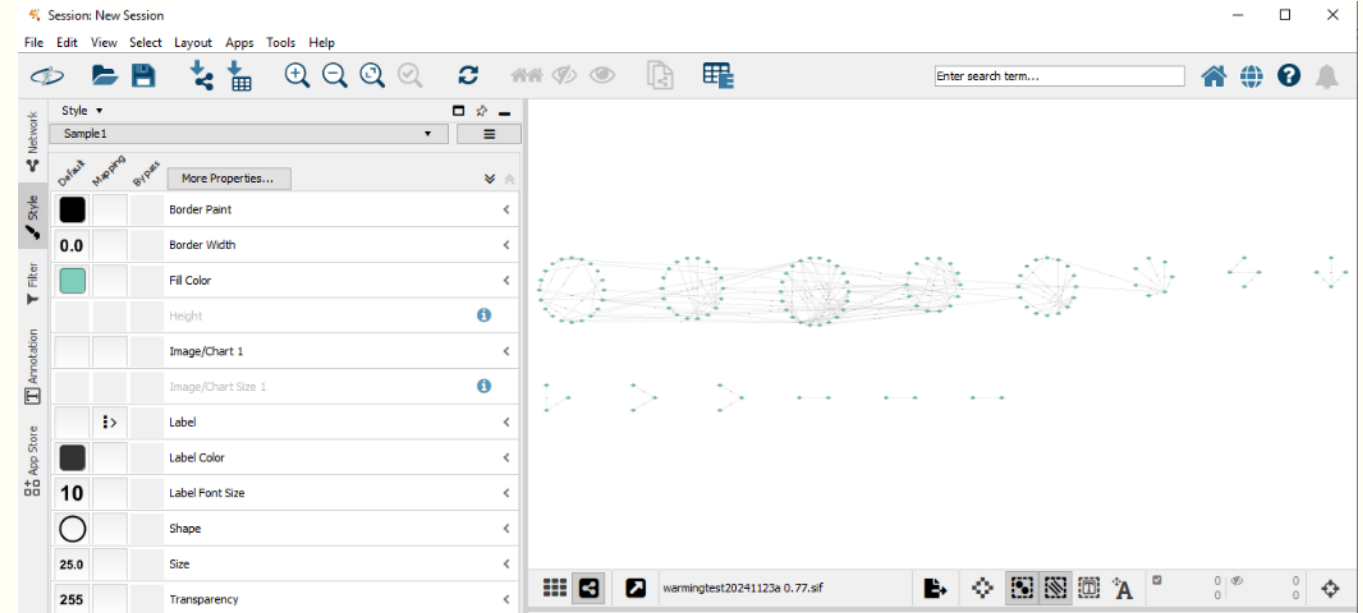
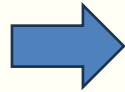
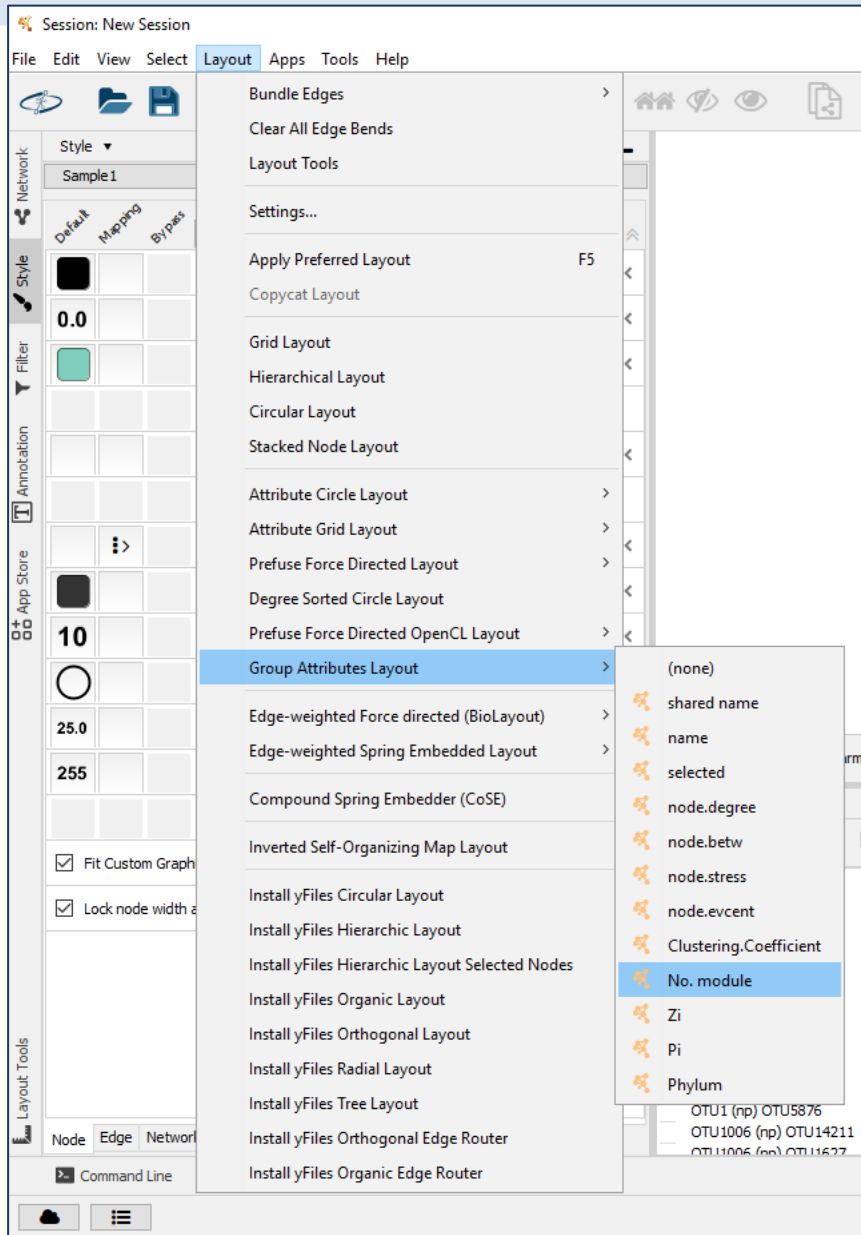
# Use Cytoscape: select style

- Select 'Style' → 'Sample 1'



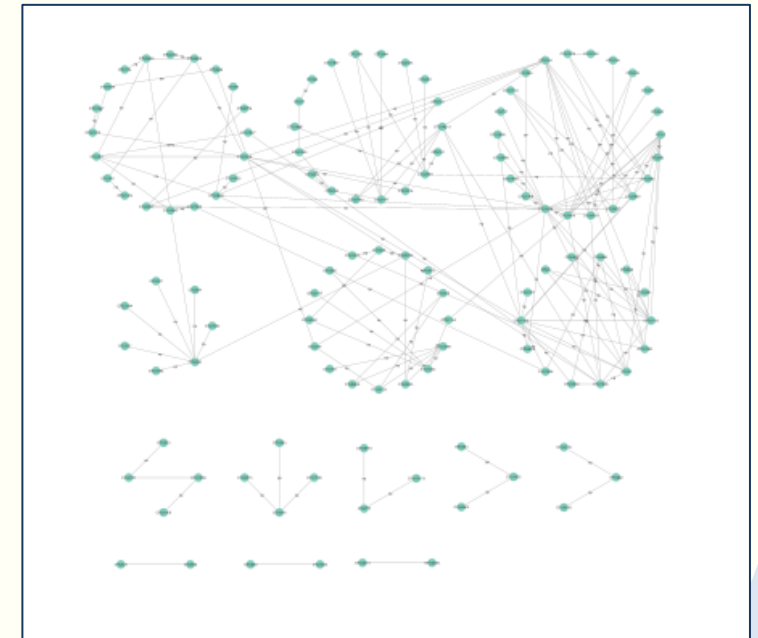
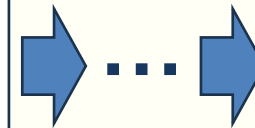
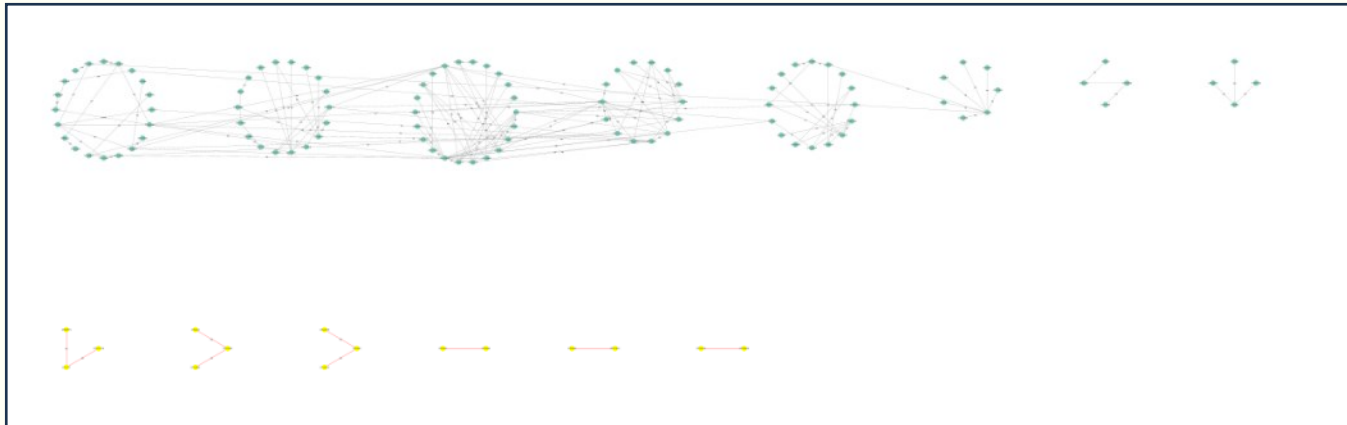
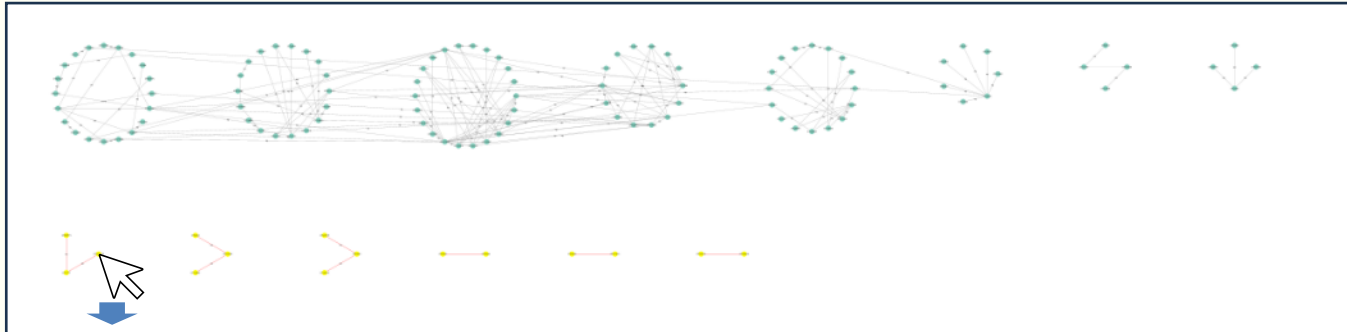
# Use Cytoscape: layout

- You may explore different layout options
- Here the example shows 'group by module'



# Use Cytoscape: manual adjustment of layout

- To select nodes and edges, hold 'Ctrl' and use mouse drag (hold down left mouse button and drag) to select. Selected nodes will become yellow, and selected edges will become red.
- Move mouse above one of the nodes you want to move, then hold down left mouse and move.
- Click any other position will deselect the nodes and edges.
- Repeatedly select nodes/edges and move to adjust the layout.



# Use Cytoscape: Node color

- You may explore node color by other different attributes.
- Here the example shows node color by phylum information.

1

2

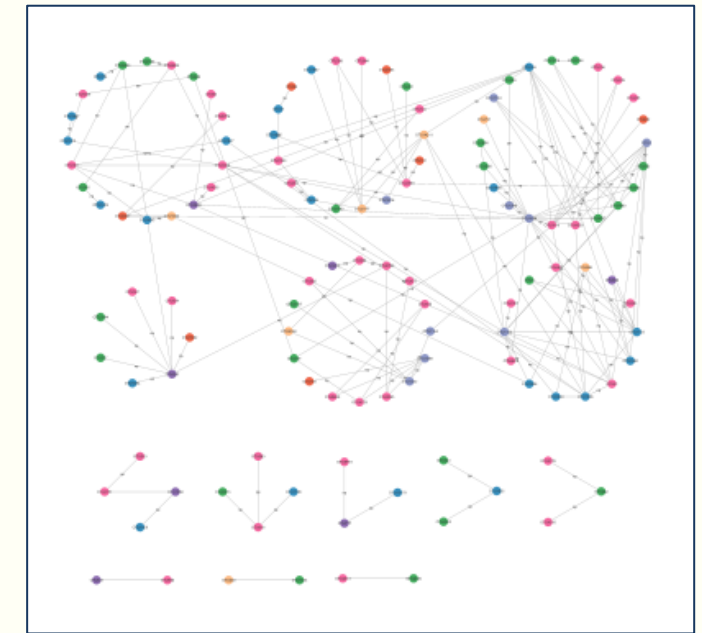
3

4

5

6

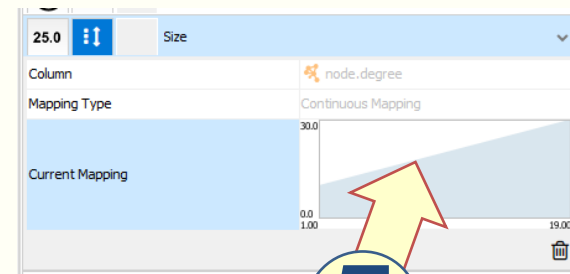
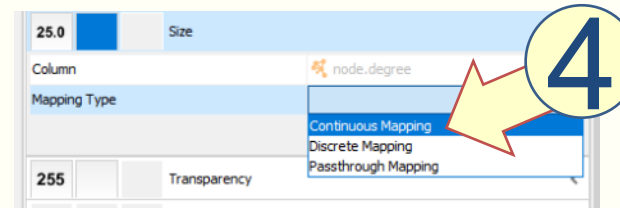
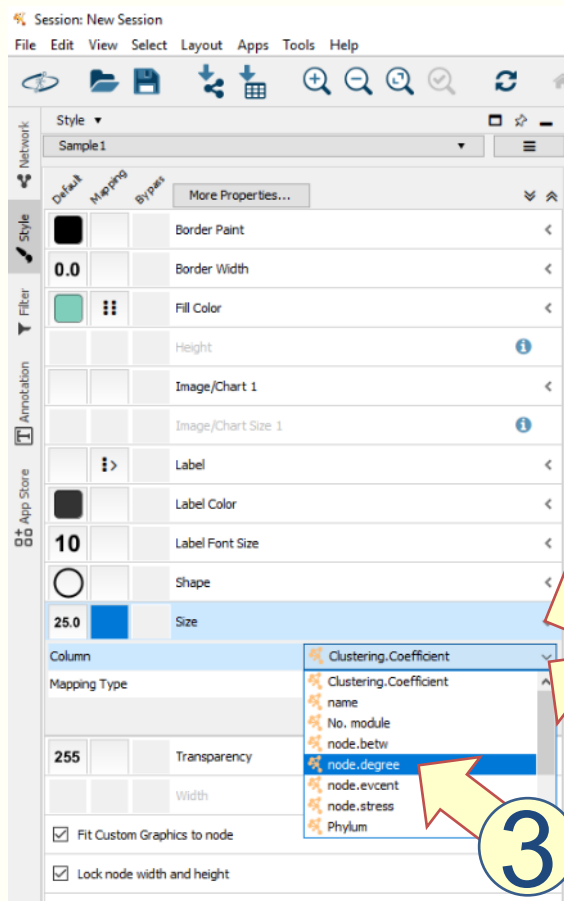
Column	Phylum
Mapping Type	Discrete Mapping
Actinobacteria	R:65 G:171 B:93 - #41AB5D
Bacteroidetes	R:54 G:144 B:192 - #3690C0
Cyanobacteria	R:239 G:101 B:72 - #EF6548
Firmicutes	R:140 G:150 B:198 - #8C96C6
Fusobacteria	R:140 G:107 B:177 - #8C6BB1
Proteobacteria	R:247 G:104 B:161 - #F768A1
Verrucomicrobia	R:253 G:187 B:132 - #FDBB84





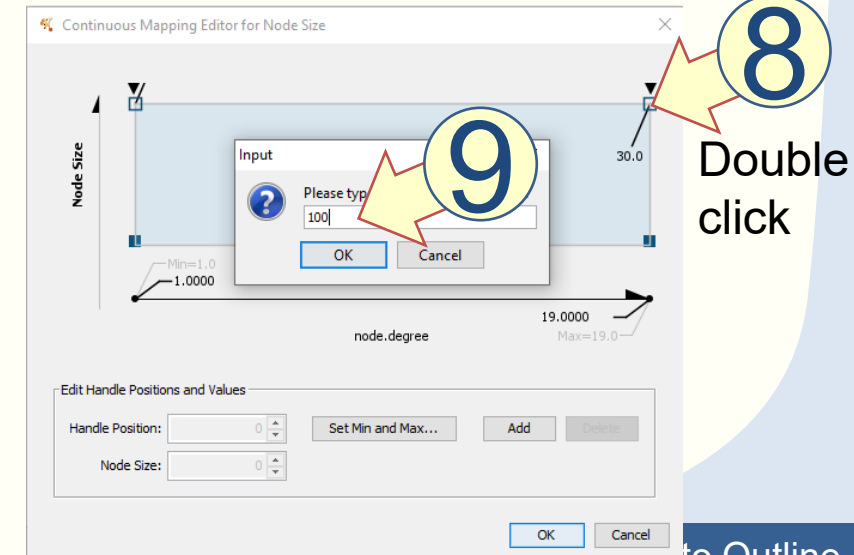
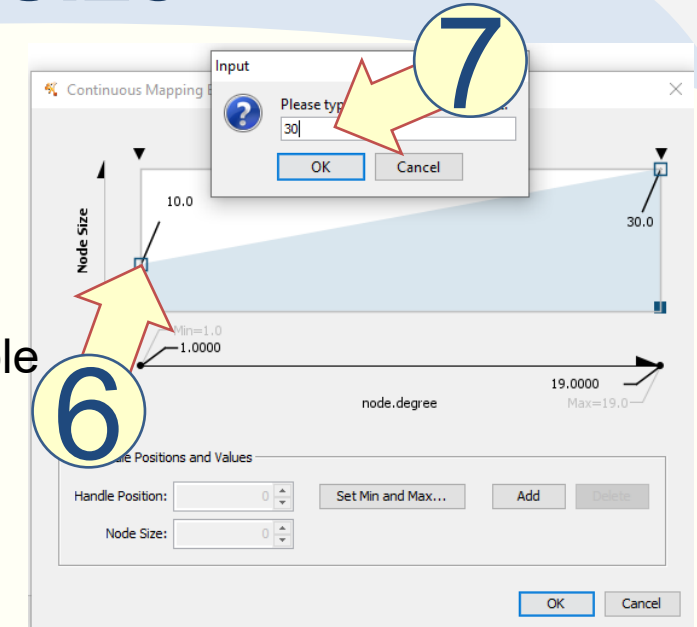
# Use Cytoscape: Node size

- You may explore node size by other attributes.
- Here the example shows node size by node degree.



Double click

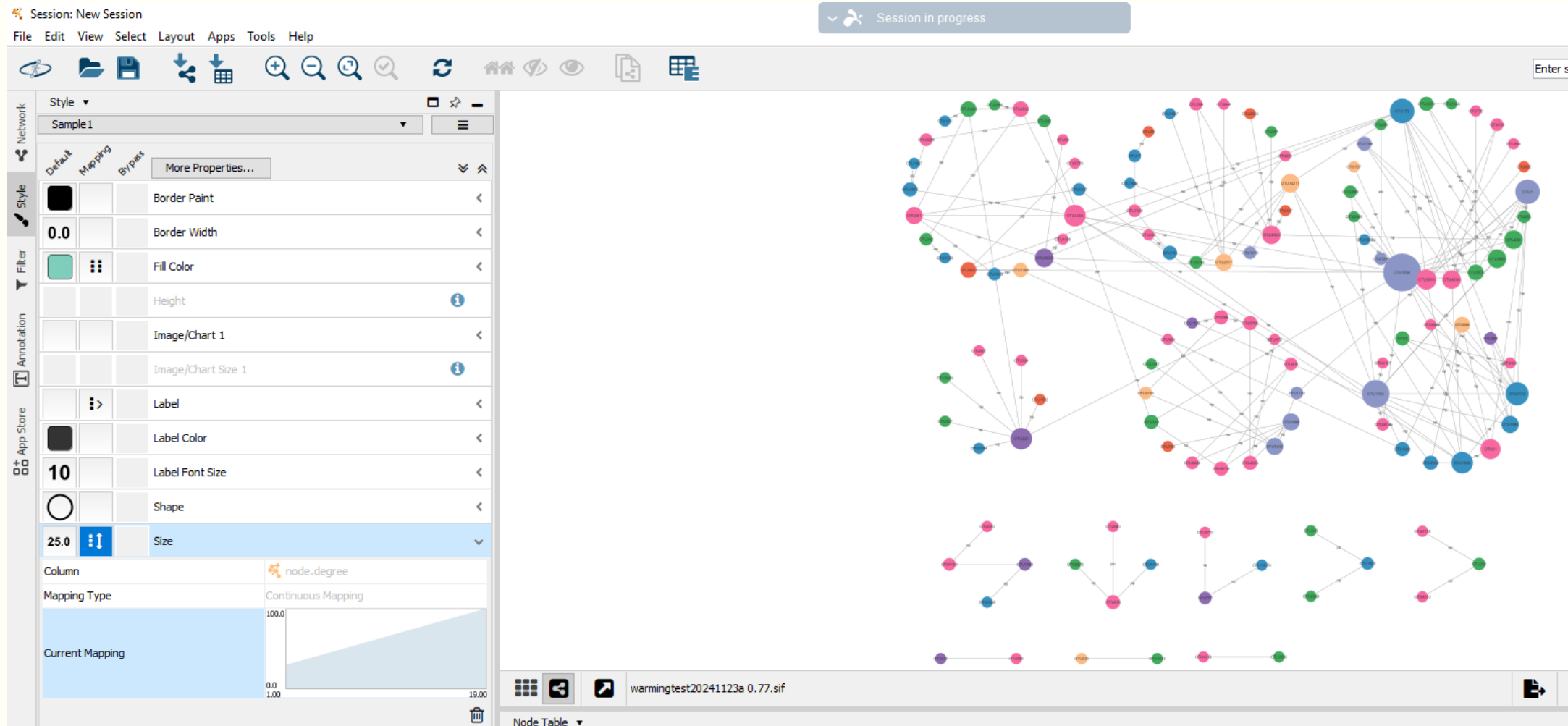
Double click



Double click

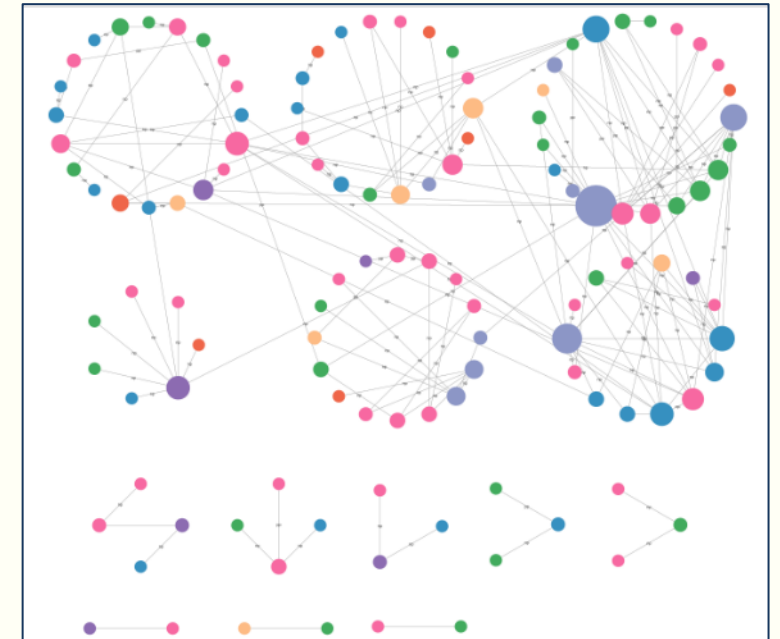
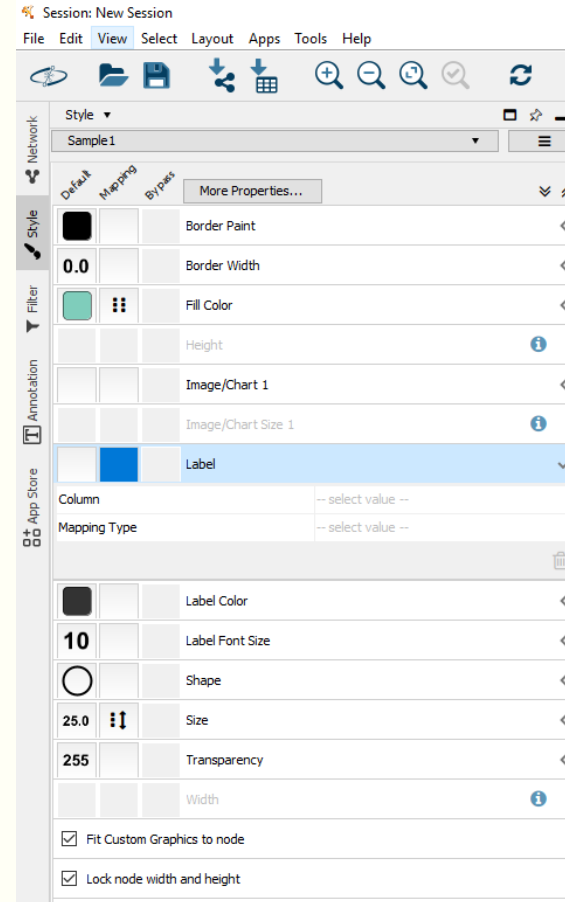
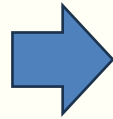
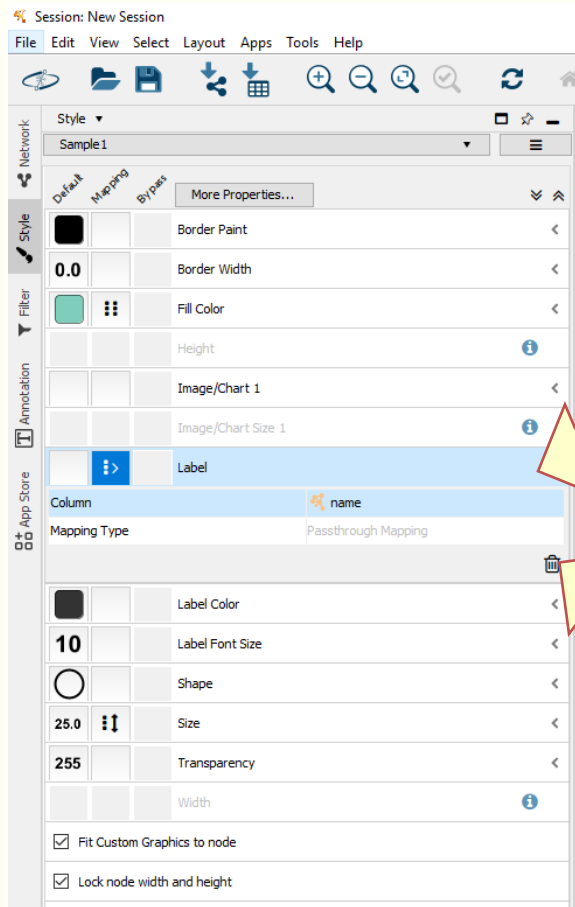
# Use Cytoscape: Node size

## Node size setting result



# Use Cytoscape: Remove node labels

- Usually, the node labels are not necessary.

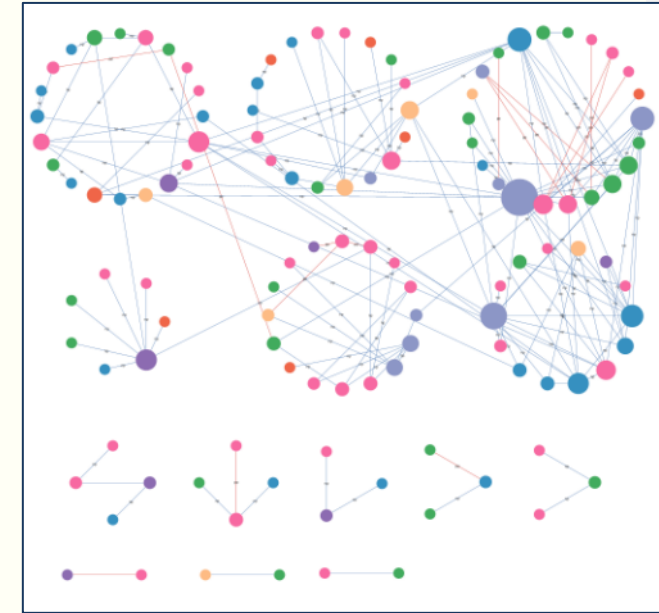


# Use Cytoscape: Edge color

- You may explore edge color by other attributes.
- Here the example shows edge color by positive vs negative correlation.

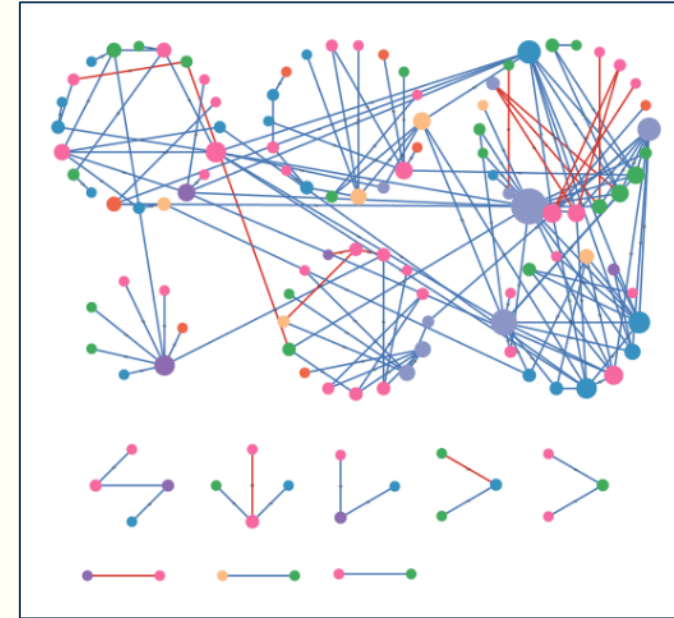
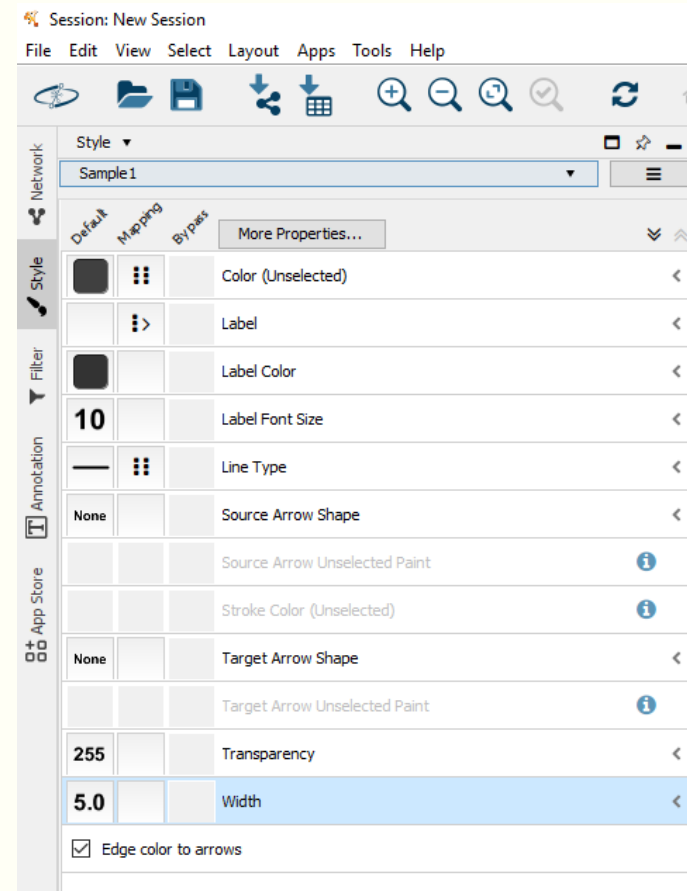
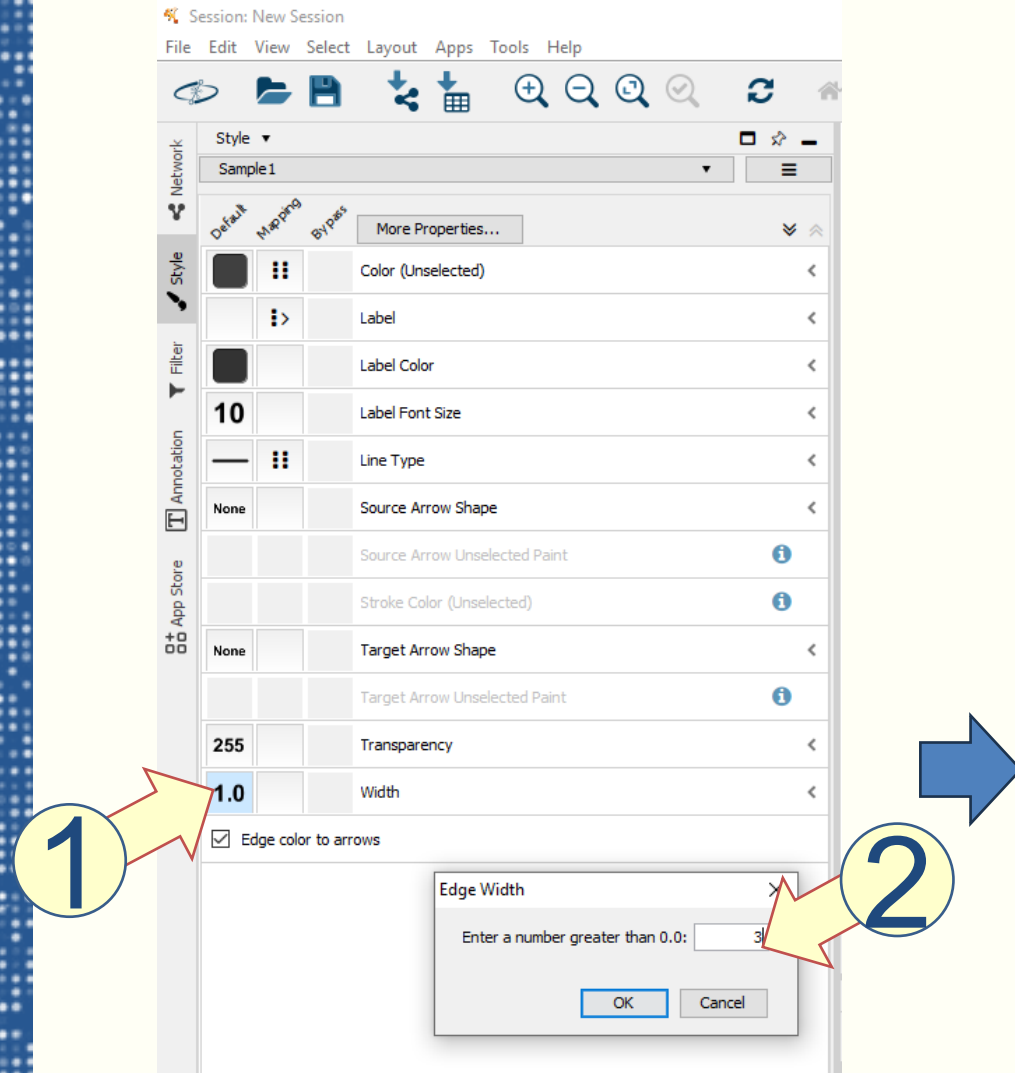
The following steps are shown in the Cytoscape interface:

1. Select 'Edge' in the 'Layout Tools' panel.
2. Select 'Color' in the 'Style' panel.
3. Select 'Column 2' in the 'Mapping Type' dropdown.
4. Select 'Discrete Mapping' in the 'Mapping Type' dropdown.
5. Select 'Discrete Mapping' in the 'Mapping Type' dropdown.
6. Map the values -1.0 (blue) and 1.0 (red) to the edge color.



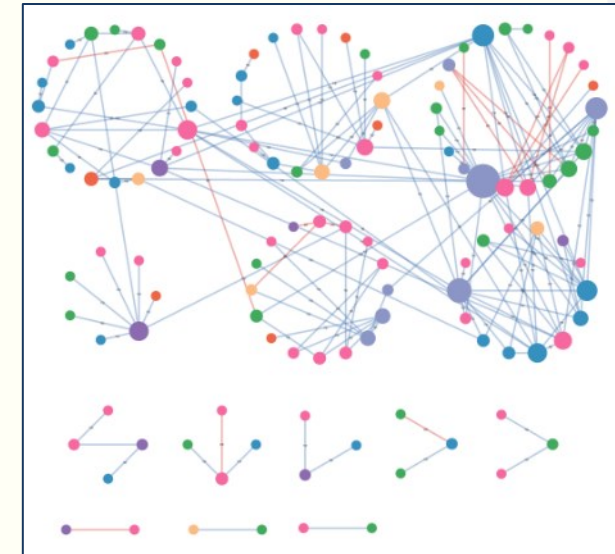
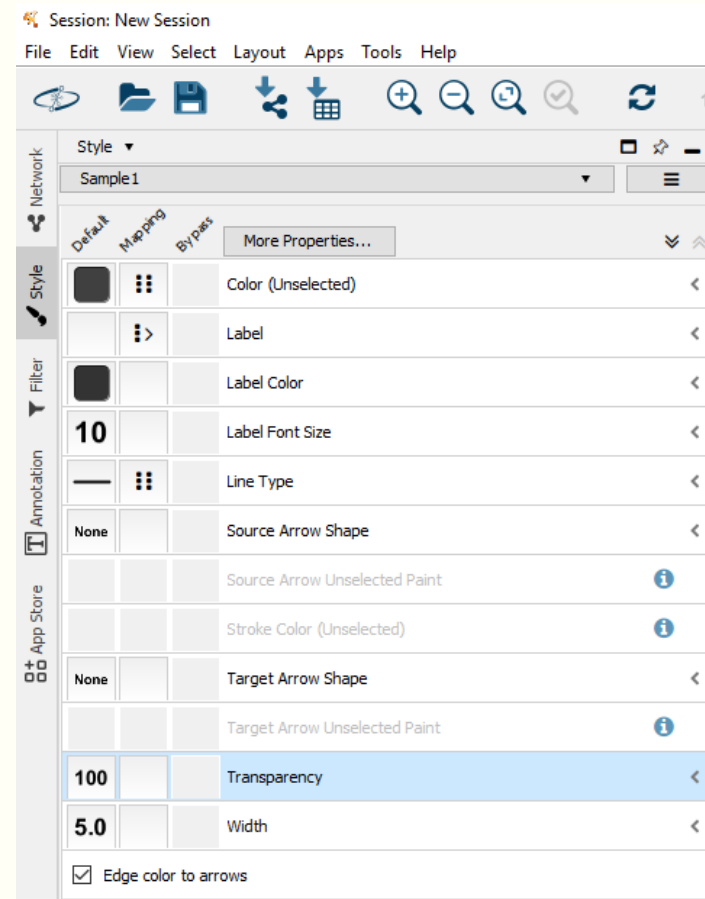
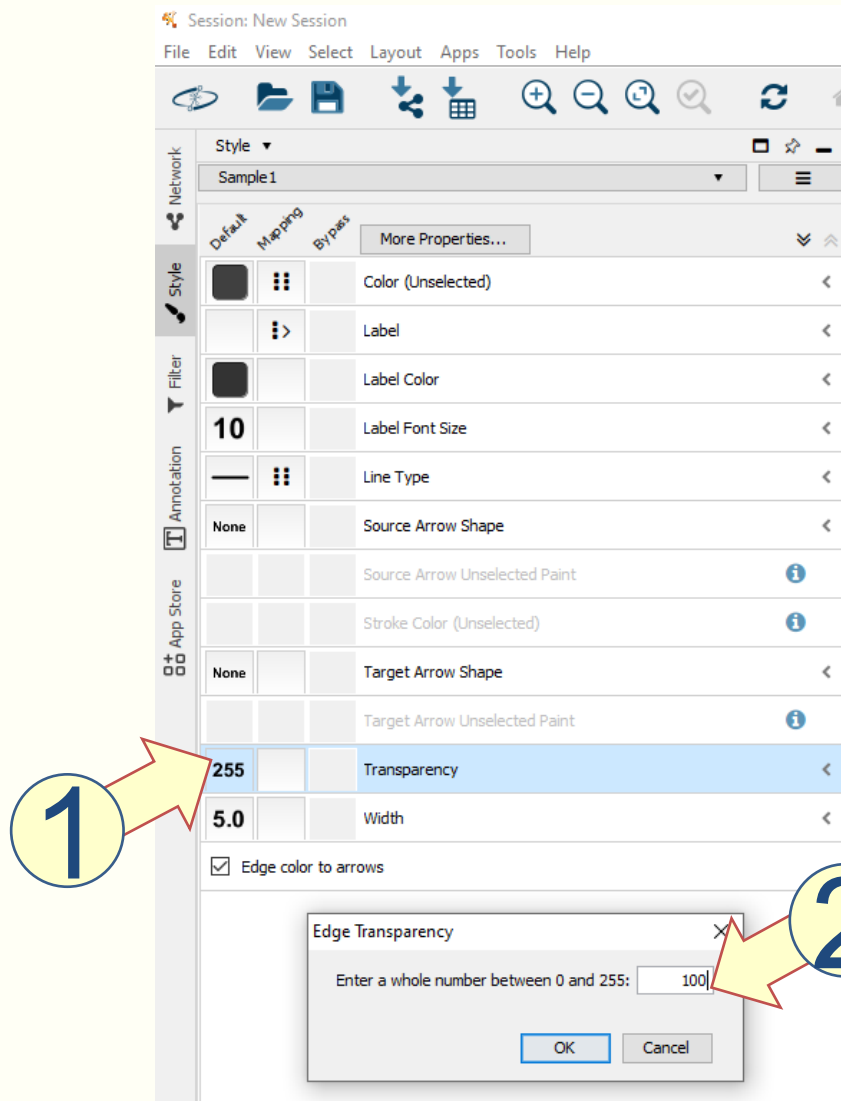


# Use Cytoscape: Edge size



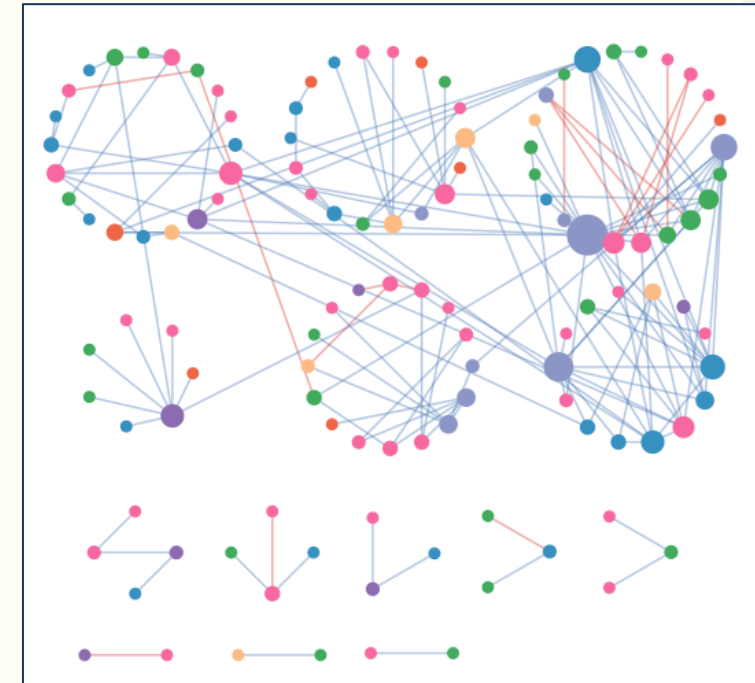
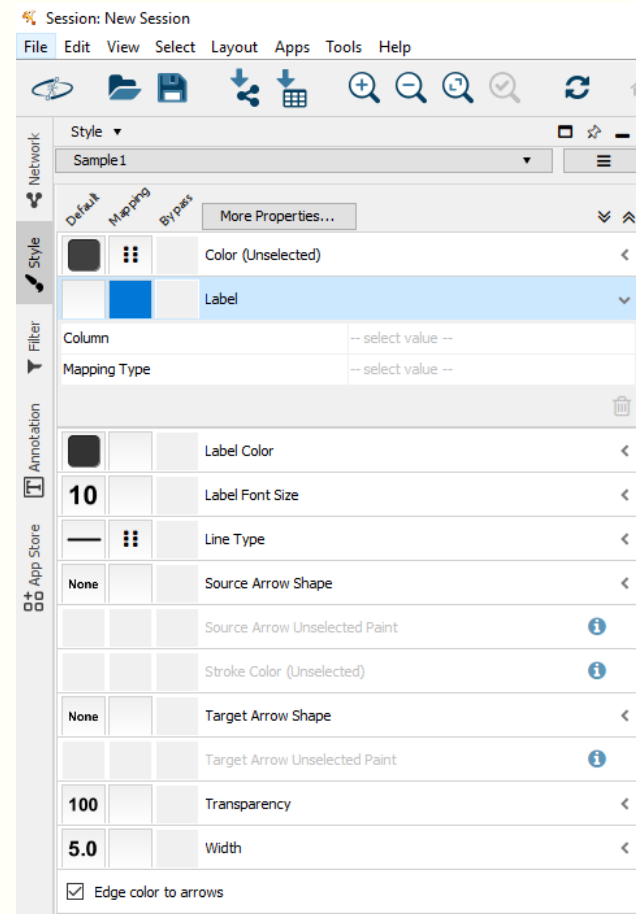
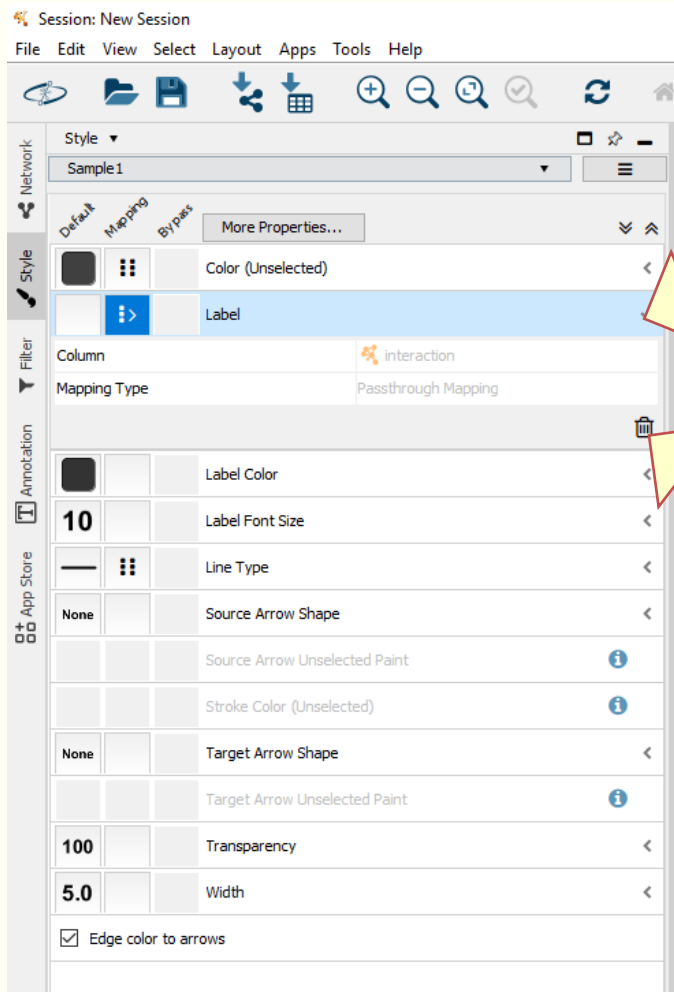
# Use Cytoscape: Edge transparency

- If you want to make the edge color lighter, you can decrease the value for transparency

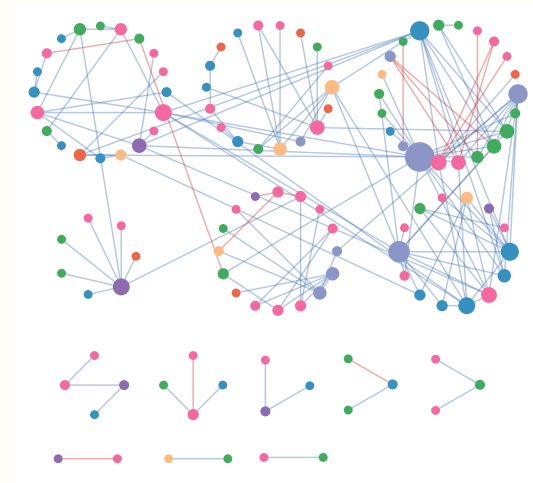
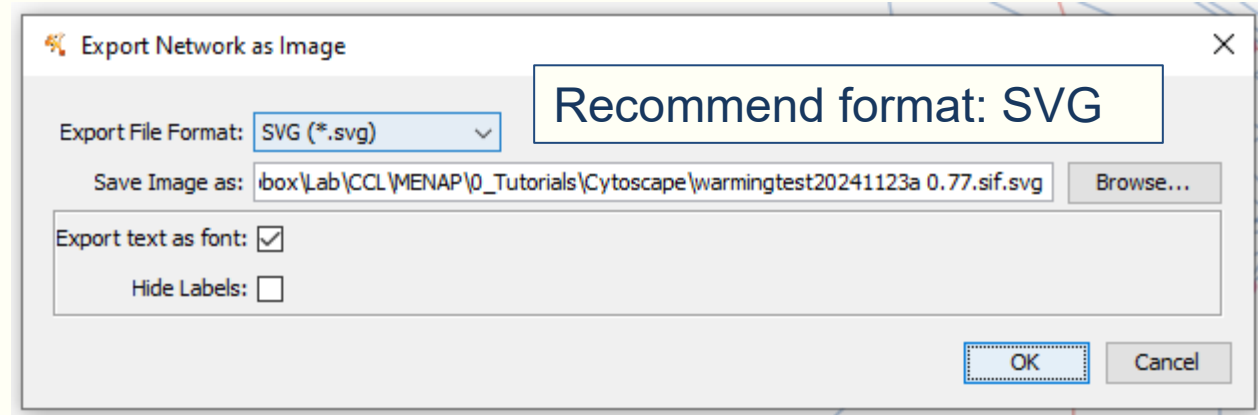
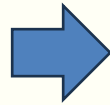
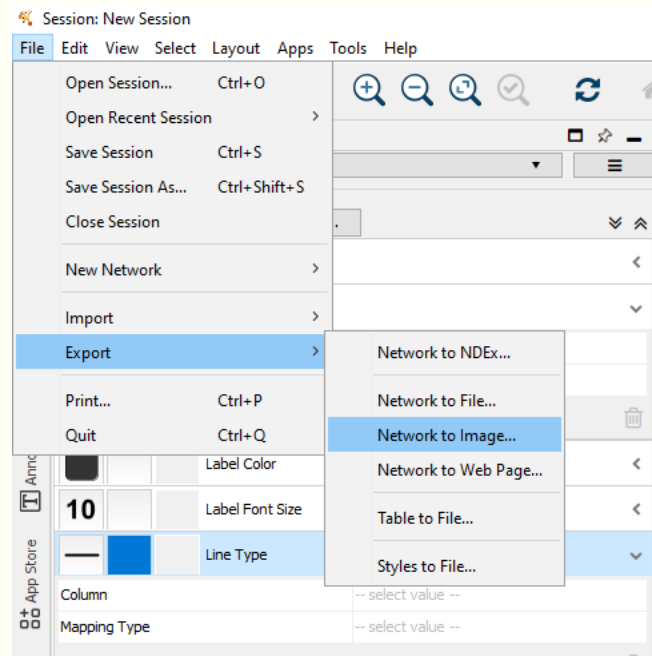


# Use Cytoscape: Remove edge label

- Usually, the edge labels are not necessary.

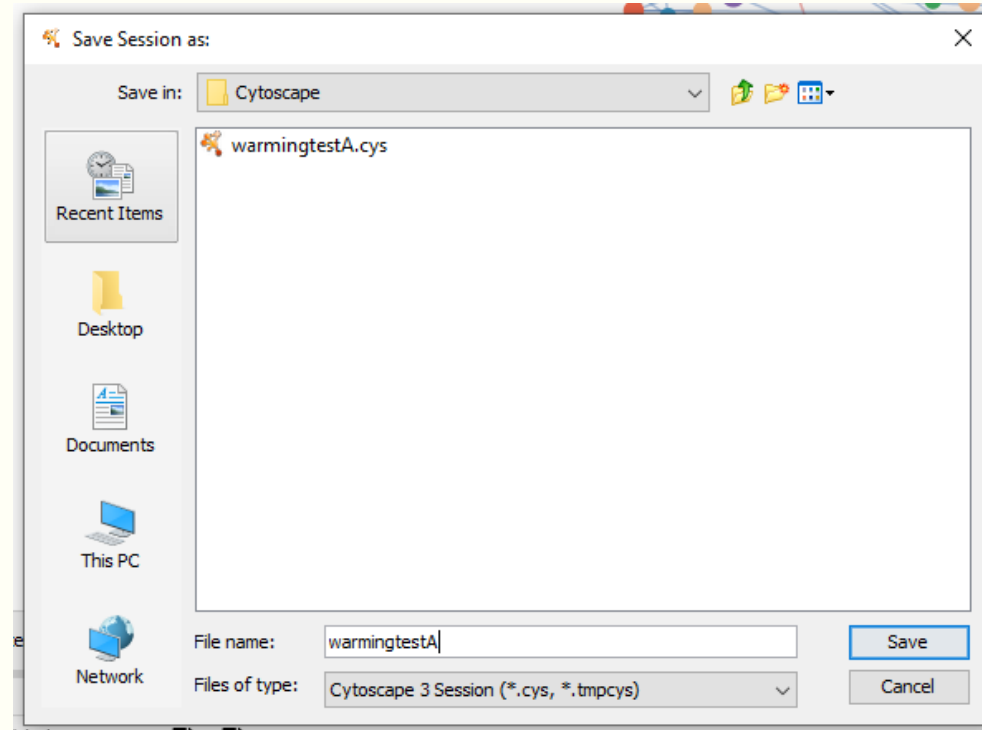
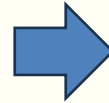
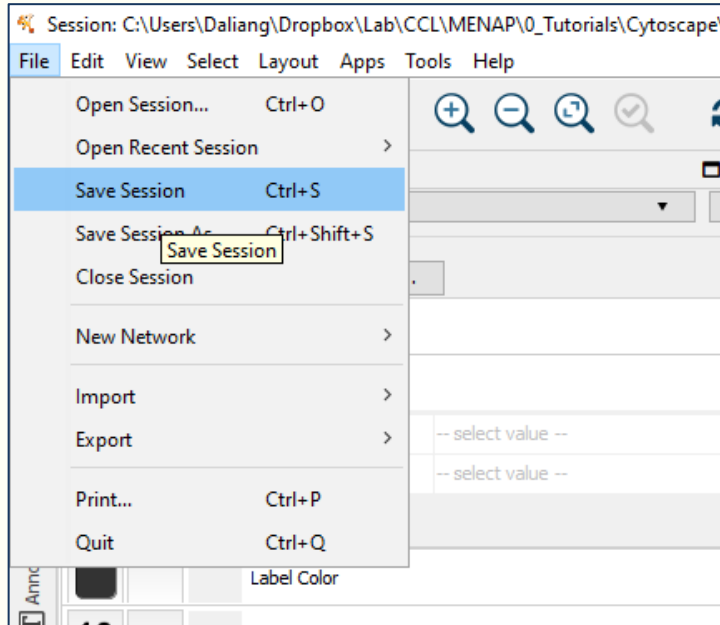


# Use Cytoscape: Export image









# Use Cytoscape: Save session



# Fit three power-law models

## Network statistics

- 1  Fit three power-law models (regular power law, exponential law and truncated power law)
-  Randomize the network structure and then calculate network properties  
(Updated: Modularity values from random networks have been added.)
-  Calculate Gene/OTU significances (GS) with environmental traits and then use Mantel test to check the correlations between GS and network connectivity
-  Module-EigenGene analyses



## Main -> Analysis

### Select Network to do power\_law\_fitting

search by:  date

order by:  date  descending  submit

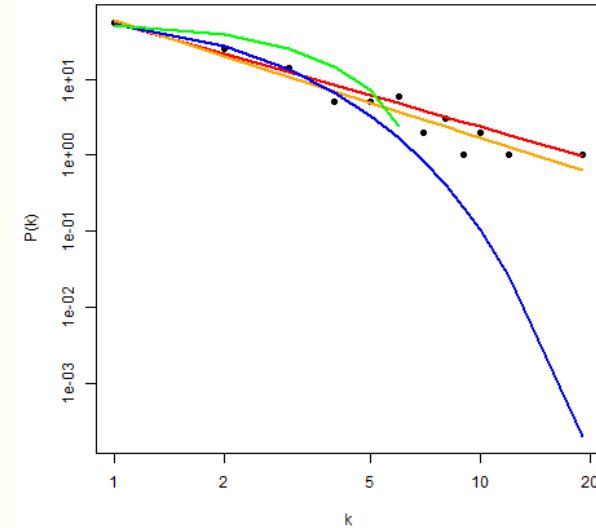
Please select a Molecular Ecological Network (MEN)

	User	Experiment name	Construction time	RMT threshold	# nodes	# links
<input checked="" type="radio"/>	Daliang	warmingtest20241123a	2024-11-23 11:29:18	0.7700	120	163
<input type="radio"/>	Daliang	M120d20240723a	2024-07-23 18:26:40	0.9000	136	110

Submit

## Main -> Analysis

### Power Law models fitting



Power-law: lamda = 1.370054, R2 of power fitting= 0.9867646  
 Log Power-law: lamda = 1.537721, R2 of log(y)-log(x): 0.9241948  
 Exponential distribution: lamda = 0.6959439, R2 = 0.9841367  
 Truncated Power-law: lamda = 0.6807769, kx = 1.616316, R2 = 0.7798388

You can [download](#) the values

Copy paste  
this part for  
your record.

Right-click 'download' and 'Save link as ...' to download the table.

# Randomize the network

## Network statistics



Fit three power-law models (regular power law, exponential law and truncated power law)



Randomize the network structure and then calculate network properties  
(Updated: Modularity values from random networks have been added.)



Calculate Gene/OTU significances (GS) with environmental traits and then use Mantel test to check the correlations between GS and network connectivity



Module-EigenGene analyses



## Main -> Analysis

### Select Network to do Randomization

search by:  date   
order by:  date  descending  submit

Please select a Molecular Ecological Network (MEN)

	User	Experiment name	Construction time	RMT threshold	# nodes	# links
<input checked="" type="radio"/>	Daliang	warmingtest20241123a	2024-11-23 11:29:18	0.7700	120	163
<input type="radio"/>	Daliang	M120d20240723a	2024-07-23 18:26:40	0.9000	136	110

Submit

## Main -> Analysis

### Randomly rewire the network connections and calculate the network properties

This is your first time to run the randomization process. It will take a while. Please come back or refresh this page to check the status. After it finishes, the results will be shown in this page.

Wait until you can refresh the page



## Main -> Analysis

### Randomly rewire the network connections and calculate the network properties

Network Indexes	Empirical Network Indexes	100 Random Networks Indexes
Average clustering coefficient (avgCC)	0.001	0.027 +/- 0.011
Average path distance (GD)	4.558	3.960 +/- 0.128
Geodesic efficiency (E)	0.278	0.296 +/- 0.007
Harmonic geodesic distance (HD)	3.602	3.375 +/- 0.079
Centralization of degree (CD)	0.139	0.139 +/- 0.000
Centralization of betweenness (CB)	0.193	0.264 +/- 0.037
Centralization of stress centrality (CS)	1.434	0.632 +/- 0.120
Centralization of eigenvector centrality (CE)	0.369	0.398 +/- 0.032
Density (D)	0.023	0.023 +/- 0.000
Reciprocity	1	1.000 +/- 0.000
Transitivity (Trans)	0.004	0.038 +/- 0.012
Connectedness (Con)	0.655	0.827 +/- 0.059
Efficiency	0.977	0.982 +/- 0.001
Hierarchy	0	0.000 +/- 0.000
Lubness	1	1.000 +/- 0.000
Modularity(fast_greedy)	0.616	0.597 +/- 0.011

Copy paste the table to a spreadsheet

# Relationship with environmental traits (1)

## Network statistics



Fit three power-law models (regular power law, exponential law and truncated power law)



Randomize the network structure and then calculate network properties  
(Updated: Modularity values from random networks have been added.)



Calculate Gene/OTU significances (GS) with environmental traits and then use Mantel test to check the correlations between GS and network connectivity

Module-EigenGene analyses

1



## Main -> Analysis

### Select MV\_dataset to do Gene\_significances

search by:

order by:

Please select a dataset to do Gene\_significances

	User	Experiment name	Upload time	File name	Number of samples	Number of genes
<input checked="" type="radio"/>	Daliang	warmingtest20241123a	2024-11-23 10:45:21	1732380321	14	228
<input type="radio"/>	Daliang	warmtest240922A	2024-09-22 18:49:48	1727048988	14	228

3

## Main -> Analysis

### Gene/OTU significances settings

Summary of your data:

Experiment name	warmingtest20241123a
# Gene/OTU	228
# Samples	14
Settings	Missing values were filled with 0.0100 by fill_paired; don't take logarithm; and then calculate Pearson Correlation.

Your environmental traits:

Please upload your file of environmental factors [here](#)

4



## Main

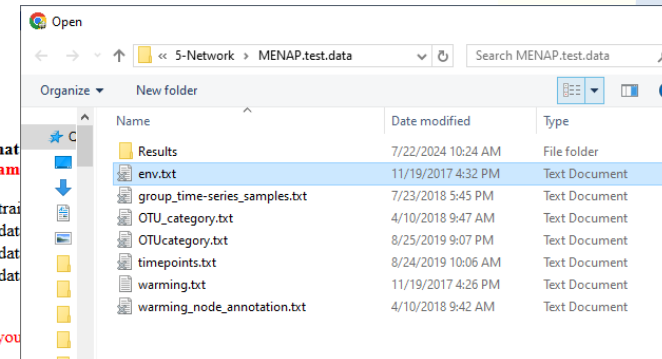
### Your experimental traits file:

No file chosen

The uploaded file must be **tab-devided text** file and its format  
The first cell (in the upper-left corner) must be **"Sample Name"**

Sample Name	trait Name1	trait Name2	trait Name3
Sample1	data11	data12	data13
Sample2	data21	data22	data23
Sample3	data31	data32	data33

Notice: please name your environmental factors as simple as you



Prepare environmental factor table according to the format requirements, then upload.

# Relationship with environmental traits (2)

Main -> Analysis

## Gene/OTU significances settings

Summary of your data:

Experiment name	warmingtest20241123a
# Gene/OTU	228
# Samples	14
Settings	Missing values were filled with 0.0100 by fill_paired; don't take logarithm; and then calculate Pearson Correlation.

Your environmental traits:

sample	pH	Moisture	TC
Warming1	3	12	30
Warming2	5	15	31
Warming3	3.2	21	32
Warming4	5.5	16	34
Warming5	6	10	34
Warming6	7	19	39
Warming7	8	18	36
Warming8	6.5	14	16
Warming9	7.1	22	12
Warming10	9	20	14
Warming11	5.4	16	19
Warming12	4.6	12	17
Warming13	3.9	11	13
Warming14	8	10	18

[Remove file](#)

Or re-upload your file of environmental traits [here](#)

Correlation method:

☒ Pearson Correlation Coefficient

☐ Do not standardize

Standardization method:

☒ standardize environmental data only (scale each factor to zero mean and unit variance)

☐ standardize genes and environmental data

☐ divide by maximum (both genes and env)

☐ divide by maximum and multiply by the number of non-zero items (both genes and env)

☐ standardize values into range 0...1 (both genes and env)

Missing values:

☐ fill with 0 (before standardization)

☐ fill with 0 (after standardization)

☒ ignore (only use paired values)

☐ fill 0 if paired with a valid value

[submit](#)

Select the options and submit.

Main -> Analysis

## Gene Significances

Notice: please wait a while and then results will appear below.

	pH	Moisture	TC
OTU1	0.211	0.082	0.091
OTU1006	0.021	0.038	0.047
OTU1012	0.002	0.005	0.296

OTU96	0.014	0.211	0.013
OTU983	0.044	0.168	0.216

You can download this table [here](#), and then open it in Excel.

[Do Mantel test](#)

Right-click 'here' and 'Save link as ...' to download the table.

Main -> Analysis

## Settings of GS vs. node connectivity

Summary of your data:

Experiment name	warmingtest20241123a
# Gene/OTU	228
# Samples	14
Gene Significance	pH,Moisture,TC

Your node annotation file:

Please upload your file with node annotations [here](#).

Distance method:

- ☒ Euclidean distance
- ☐ Bray-Cutis distance
- ☐ Morisita distance
- ☐ Jaccard distance

GS included in Mantel test:

GS	include	partial
pH	<input checked="" type="checkbox"/>	<input type="checkbox"/>
Moisture	<input checked="" type="checkbox"/>	<input type="checkbox"/>
TC	<input checked="" type="checkbox"/>	<input type="checkbox"/>

☐ Run Mantel test on each category

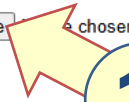

[submit](#)



# Relationship with environmental traits (3)

**Main**

**Your node annotation file:**

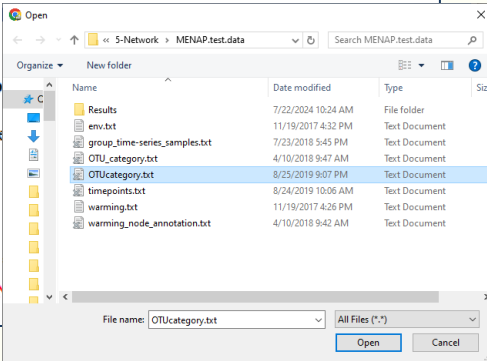
Choose File  

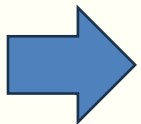
Submit

The uploaded file must be a **tab-separated text file** and its format should be as follows:

Node ID	Gene_name1/Phylogeny1	Gene_name2/Phylogeny2
ID1	data11	data12
ID2	data21	data22
ID3	data31	data32
...		

Notice: please name your annotation names as simple as you can and DO NOT use special characters.





**Main -> Analysis**

**Settings of GS vs. node connectivity**

Summary of your data:

Experiment name	warmingtest20241123a
# Gene/OTU	228
# Samples	14
Gene Significance	pH,Moisture,TC

**Your node annotation file:**



# nodes	976
Category	Phylum

Notice: it seems your uploaded node annotation file has unequal number of nodes than your network included.

[Remove file](#)

Or re-upload your file of environmental traits [here](#)

**Distance method:**

☒ Euclidean distance  


☐ Bray-Cutis distance

☐ Morisita distance

☐ Jaccard distance



**GS included in Mantel test:**



GS	include	partial
pH	<input checked="" type="checkbox"/>	<input type="checkbox"/>
Moisture	<input checked="" type="checkbox"/>	<input type="checkbox"/>
TC	<input checked="" type="checkbox"/>	<input type="checkbox"/>

 **Select as you need**

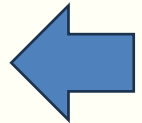
☒ Run Mantel test on each category

**Run category on:**

☒ Phylum   **Select as you need**

Submit



**Main -> Analysis**

**Mantel test (correlation) on GS vs. node connectivity**

	r	p
Proteobacteria	-0.11535029	0.9170000
Acidobacteria	-0.18522427	0.9590000
Planctomycetes	0.13182372	0.1420000
Firmicutes	-0.09376683	0.8390000
Bacteroidetes	-0.28608614	0.8916667
Actinobacteria	-0.03194668	0.5000000
Omnitrophica	0.93467553	0.2000000
Chloroflexi	0.88157740	0.0110000

# Module-EigenGene analyses (1)

## Network statistics



Fit three power-law models (regular power law, exponential law and truncated power law)



Randomize the network structure and then calculate network properties  
(Updated: Modularity values from random networks have been added.)



Calculate Gene/OTU significances (GS) with environmental traits and then use Mantel test to check the correlations between GS and network connectivity

## Module-EigenGene analyses

1



## Main -> Analysis

### Select MV\_dataset to do Module-EigenGene

search by:

order by:

Please select a dataset to do Module-EigenGene

	User	Experiment name	Upload time	File name	Number of samples	Number of genes
<input checked="" type="radio"/>	Daliang	warmingtest20241123a	2024-11-23 10:45:21	1732380321	14	228
<input type="radio"/>	Daliang	warmtest240922A	2024-09-22 18:49:48	1727048988	14	228

3

## Main -> Analysis

### Module-EigenGene settings

Summary of your uploaded data:

Experiment name	warmingtest20241123a
# Gene/OTU	228
# Samples	14
Settings	Missing values were filled with 0.0100 by fill_paired; don't take logarithm; and then calculate Pearson Correlation.

Your environmental traits:

Please upload your file of environmental factors [here](#).

4

## Main

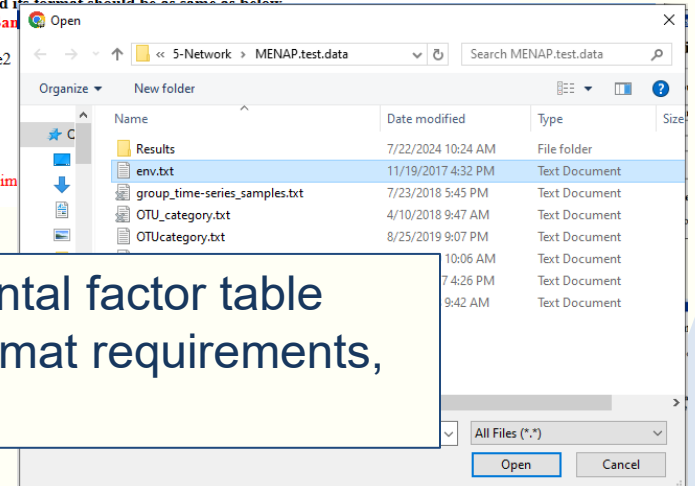
Your experimental traits file:

No file chosen

The uploaded file must be **tab-devided text file** and its format should be as some as below:  
The first cell (in the upper-left corner) must be "Sample Name"

Sample Name	trait Name1	trait Name2
Sample1	data11	data12
Sample2	data21	data22
Sample3	data31	data32

Notice: please name your environmental factors as sim



Prepare environmental factor table according to the format requirements, then upload.

# Module-EigenGene analyses (2)

[Main](#) -> [Analysis](#)

## Module-EigenGene settings

Summary of your uploaded data:

Experiment name	warmingtest20241123a
# Gene/OTU	228
# Samples	14
Settings	Missing values were filled with 0.0100 by fill_paired; don't take logarithm; and then calculate Pearson Correlation.

Your environmental traits:

sample	pH	Moisture	TC
Warming1	3	12	30
Warming2	5	15	31
Warming3	3.2	21	32
Warming4	5.5	16	34
Warming5	6	10	34
Warming6	7	19	39
Warming7	8	18	36
Warming8	6.5	14	16
Warming9	7.1	22	12
Warming10	9	20	14
Warming11	5.4	16	19
Warming12	4.6	12	17
Warming13	3.9	11	13
Warming14	8	10	18

[Remove file](#)

Or re-upload your file of environmental traits [here](#)

You already ran module-separation methods on [greedy modularity optimization](#). Please choose one:

☒ greedy modularity optimization

Ignore the small modules with less than  members

[Main](#) -> [Analysis](#)

## Module-EigenGene analysis

Your first-time to run Module-eigengene analysis on this network. Record this webpage and come back to view results later or refresh this page to check the status...

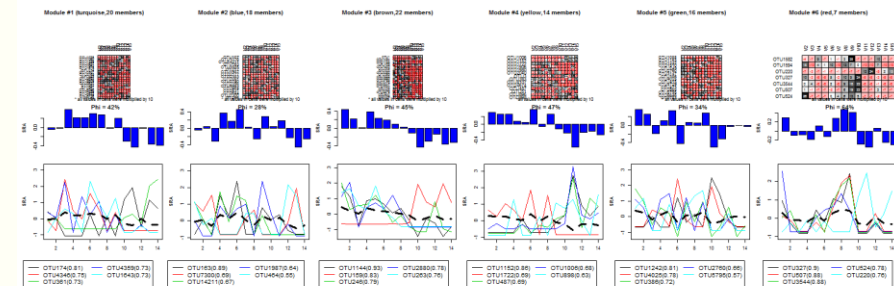
Wait until you can refresh the page

[Main](#) -> [Analysis](#)

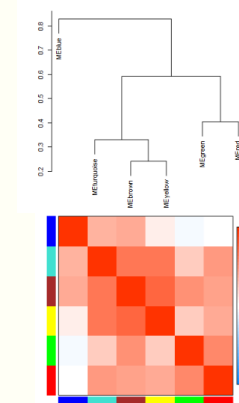
## Module-EigenGene analysis

The results of Module Eigengene analysis:

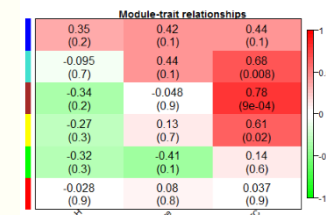
The Module-EigenGenes with module members in each module:



The Module Eigengene hierarchy structure:



The Module correlation with environmental factors:



The Module Membership table can be download [here](#)

The Module Separation table can be download [here](#)

The Module-Trait Relationship table can be download [here](#)

The Module EigenGene table can be download [here](#)

[Back to Outline](#)



**End**